

$F7\_12 = F7\_7 + F8\_8 + F9\_9 + F10\_10 + F11\_11 + F12\_12$   
 $F7\_13 = F7\_7 + F8\_8 + F9\_9 + F10\_10 + F11\_11 + F12\_12 + F13\_13$   
 $F7\_14 = F7\_7 + F8\_8 + F9\_9 + F10\_10 + F11\_11 + F12\_12 + F13\_13 + F14\_14$   
 $F7\_15 = F7\_7 + F8\_8 + F9\_9 + F10\_10 + F11\_11 + F12\_12 + F13\_13 + F14\_14 + F15\_15$   
5  $F7\_16 = F7\_7 + F8\_8 + F9\_9 + F10\_10 + F11\_11 + F12\_12 + F13\_13 + F14\_14 + F15\_15 + F16\_16$   
 $F7\_17 = F7\_7 + F8\_8 + F9\_9 + F10\_10 + F11\_11 + F12\_12 + F13\_13 + F14\_14 + F15\_15 + F16\_16 + F17\_17$   
 $F7\_18 = F7\_7 + F8\_8 + F9\_9 + F10\_10 + F11\_11 + F12\_12 + F13\_13 + F14\_14 + F15\_15 + F16\_16 + F17\_17 + F18\_18$   
 $F8\_8 = F8\_8$   
 $F8\_9 = F8\_8 + F9\_9$   
10  $F8\_10 = F8\_8 + F9\_9 + F10\_10$   
 $F8\_11 = F8\_8 + F9\_9 + F10\_10 + F11\_11$   
 $F8\_12 = F8\_8 + F9\_9 + F10\_10 + F11\_11 + F12\_12$   
 $F8\_13 = F8\_8 + F9\_9 + F10\_10 + F11\_11 + F12\_12 + F13\_13$   
 $F8\_14 = F8\_8 + F9\_9 + F10\_10 + F11\_11 + F12\_12 + F13\_13 + F14\_14$   
15  $F8\_15 = F8\_8 + F9\_9 + F10\_10 + F11\_11 + F12\_12 + F13\_13 + F14\_14 + F15\_15$   
 $F8\_16 = F8\_8 + F9\_9 + F10\_10 + F11\_11 + F12\_12 + F13\_13 + F14\_14 + F15\_15 + F16\_16$   
 $F8\_17 = F8\_8 + F9\_9 + F10\_10 + F11\_11 + F12\_12 + F13\_13 + F14\_14 + F15\_15 + F16\_16 + F17\_17$   
 $F8\_18 = F8\_8 + F9\_9 + F10\_10 + F11\_11 + F12\_12 + F13\_13 + F14\_14 + F15\_15 + F16\_16 + F17\_17 + F18\_18$   
 $F9\_9 = F9\_9$   
20  $F9\_10 = F9\_9 + F10\_10$   
 $F9\_11 = F9\_9 + F10\_10 + F11\_11$   
 $F9\_12 = F9\_9 + F10\_10 + F11\_11 + F12\_12$   
 $F9\_13 = F9\_9 + F10\_10 + F11\_11 + F12\_12 + F13\_13$   
 $F9\_14 = F9\_9 + F10\_10 + F11\_11 + F12\_12 + F13\_13 + F14\_14$   
25  $F9\_15 = F9\_9 + F10\_10 + F11\_11 + F12\_12 + F13\_13 + F14\_14 + F15\_15$   
 $F9\_16 = F9\_9 + F10\_10 + F11\_11 + F12\_12 + F13\_13 + F14\_14 + F15\_15 + F16\_16$   
 $F9\_17 = F9\_9 + F10\_10 + F11\_11 + F12\_12 + F13\_13 + F14\_14 + F15\_15 + F16\_16 + F17\_17$   
 $F9\_18 = F9\_9 + F10\_10 + F11\_11 + F12\_12 + F13\_13 + F14\_14 + F15\_15 + F16\_16 + F17\_17 + F18\_18$   
 $F10\_10 = F10\_10$   
30  $F10\_11 = F10\_10 + F11\_11$   
 $F10\_12 = F10\_10 + F11\_11 + F12\_12$   
 $F10\_13 = F10\_10 + F11\_11 + F12\_12 + F13\_13$   
 $F10\_14 = F10\_10 + F11\_11 + F12\_12 + F13\_13 + F14\_14$   
 $F10\_15 = F10\_10 + F11\_11 + F12\_12 + F13\_13 + F14\_14 + F15\_15$   
35  $F10\_16 = F10\_10 + F11\_11 + F12\_12 + F13\_13 + F14\_14 + F15\_15 + F16\_16$   
 $F10\_17 = F10\_10 + F11\_11 + F12\_12 + F13\_13 + F14\_14 + F15\_15 + F16\_16 + F17\_17$   
 $F10\_18 = F10\_10 + F11\_11 + F12\_12 + F13\_13 + F14\_14 + F15\_15 + F16\_16 + F17\_17 + F18\_18$   
 $F11\_11 = F11\_11$   
 $F11\_12 = F11\_11 + F12\_12$   
40  $F11\_13 = F11\_11 + F12\_12 + F13\_13$   
 $F11\_14 = F11\_11 + F12\_12 + F13\_13 + F14\_14$   
 $F11\_15 = F11\_11 + F12\_12 + F13\_13 + F14\_14 + F15\_15$   
 $F11\_16 = F11\_11 + F12\_12 + F13\_13 + F14\_14 + F15\_15 + F16\_16$   
 $F11\_17 = F11\_11 + F12\_12 + F13\_13 + F14\_14 + F15\_15 + F16\_16 + F17\_17$   
45  $F11\_18 = F11\_11 + F12\_12 + F13\_13 + F14\_14 + F15\_15 + F16\_16 + F17\_17 + F18\_18$   
 $F12\_12 = F12\_12$   
 $F12\_13 = F12\_12 + F13\_13$   
 $F12\_14 = F12\_12 + F13\_13 + F14\_14$   
 $F12\_15 = F12\_12 + F13\_13 + F14\_14 + F15\_15$   
50  $F12\_16 = F12\_12 + F13\_13 + F14\_14 + F15\_15 + F16\_16$   
 $F12\_17 = F12\_12 + F13\_13 + F14\_14 + F15\_15 + F16\_16 + F17\_17$   
 $F12\_18 = F12\_12 + F13\_13 + F14\_14 + F15\_15 + F16\_16 + F17\_17 + F18\_18$   
 $F13\_13 = F13\_13$   
 $F13\_14 = F13\_13 + F14\_14$   
55  $F13\_15 = F13\_13 + F14\_14 + F15\_15$   
 $F13\_16 = F13\_13 + F14\_14 + F15\_15 + F16\_16$   
 $F13\_17 = F13\_13 + F14\_14 + F15\_15 + F16\_16 + F17\_17$   
 $F13\_18 = F13\_13 + F14\_14 + F15\_15 + F16\_16 + F17\_17 + F18\_18$   
 $F14\_14 = F14\_14$   
60  $F14\_15 = F14\_14 + F15\_15$   
 $F14\_16 = F14\_14 + F15\_15 + F16\_16$   
 $F14\_17 = F14\_14 + F15\_15 + F16\_16 + F17\_17$   
 $F14\_18 = F14\_14 + F15\_15 + F16\_16 + F17\_17 + F18\_18$

F15\_15 = F15\_15  
 F15\_16 = F15\_15 + F16\_16  
 F15\_17 = F15\_15 + F16\_16 + F17\_17  
 F15\_18 = F15\_15 + F16\_16 + F17\_17 + F18\_18  
 F16\_16 = F16\_16  
 F16\_17 = F16\_16 + F17\_17  
 F16\_18 = F16\_16 + F17\_17 + F18\_18  
 F17\_17 = F17\_17  
 F17\_18 = F17\_17 + F18\_18  
 F18\_18 = F18\_18

Once the sequence of each pre-ligated fragment is determined, the system begins to estimate the portions of each pre-ligated sequence to be used to generate the desired PDF. As discussed above, the ligation reaction for a sequence having 18 fragments preferably takes place as 18 separate reactions. Thus, the system generates a starting set of ligation reactions for each of the 18 separate ligations. It should be noted that each ligation step uses progressively fewer of the pre-ligated molecules. This is due to the fact that, for example, the third step of the ligation reaction would not require pre-ligated fragments starting with fragment 1 "F1" or fragment 2 (F2) since these fragments have already been ligated to other fragments by the third step in the ligation. At step three, there should only ligation of fragments that bind to the third fragment from each parent.

For example, the following are exemplary ligation reactions that take place within the memory of the computer system.

Number of Ligation Steps: 18

Simulated Ligation volume of each step (ul): 100

Ligation Step #1	Ligation Step #2	Ligation Step #3	Ligation Step #4	Ligation Step #5
0.6 ul of F1_1	0.7 ul of F2_2	0.7 ul of F3_3	0.8 ul of F4_4	1.0 ul of F5_5
1.2 ul of F1_2	1.3 ul of F2_3	1.5 ul of F3_4	1.7 ul of F4_5	1.9 ul of F5_6
1.8 ul of F1_3	2.0 ul of F2_4	2.2 ul of F3_5	2.5 ul of F4_6	2.9 ul of F5_7
2.3 ul of F1_4	2.6 ul of F2_5	2.9 ul of F3_6	3.3 ul of F4_7	3.8 ul of F5_8
2.9 ul of F1_5	3.3 ul of F2_6	3.7 ul of F3_7	4.2 ul of F4_8	4.8 ul of F5_9
3.5 ul of F1_6	3.9 ul of F2_7	4.4 ul of F3_8	5.0 ul of F4_9	5.7 ul of F5_10
4.1 ul of F1_7	4.6 ul of F2_8	5.1 ul of F3_9	5.8 ul of F4_10	6.7 ul of F5_11
4.7 ul of F1_8	5.2 ul of F2_9	5.9 ul of F3_10	6.7 ul of F4_11	7.6 ul of F5_12
5.3 ul of F1_9	5.9 ul of F2_10	6.6 ul of F3_11	7.5 ul of F4_12	8.6 ul of F5_13
5.8 ul of F1_10	6.5 ul of F2_11	7.4 ul of F3_12	8.3 ul of F4_13	9.5 ul of F5_14
6.4 ul of F1_11	7.2 ul of F2_12	8.1 ul of F3_13	9.2 ul of F4_14	10.5 ul of F5_15

7.0 ul of F1_12 7.6 ul of F1_13 8.2 ul of F1_14 8.8 ul of F1_15 9.4 ul of F1_16 9.9 ul of F1_17 10.5 ul of F1_18	7.8 ul of F2_13 8.5 ul of F2_14 9.2 ul of F2_15 9.8 ul of F2_16 10.5 ul of F2_17 11.1 ul of F2_18	8.8 ul of F3_14 9.6 ul of F3_15 10.3 ul of F3_16 11.0 ul of F3_17 11.8 ul of F3_18	10.0 ul of F4_15 10.8 ul of F4_16 11.7 ul of F4_17 12.5 ul of F4_18	11.4 ul of F5_16 12.4 ul of F5_17 13.3 ul of F5_18
<b>Ligation Step #6:</b>	<b>Ligation Step #7</b>	<b>Ligation Step #8</b>	<b>Ligation Step #9</b>	<b>Ligation Step #10</b>
1.1 ul of F6_6 2.2 ul of F6_7 3.3 ul of F6_8 4.4 ul of F6_9 5.5 ul of F6_10 6.6 ul of F6_11 7.7 ul of F6_12 8.8 ul of F6_13 9.9 ul of F6_14 11.0 ul of F6_15 12.1 ul of F6_16 13.2 ul of F6_17 14.3 ul of F6_18	1.3 ul of F7_7 2.6 ul of F7_8 3.8 ul of F7_9 5.1 ul of F7_10 6.4 ul of F7_11 7.7 ul of F7_12 9.0 ul of F7_13 10.3 ul of F7_14 11.5 ul of F7_15 12.8 ul of F7_16 14.1 ul of F7_17 15.4 ul of F7_18	1.5 ul of F8_8 3.0 ul of F8_9 4.5 ul of F8_10 6.1 ul of F8_11 7.6 ul of F8_12 9.1 ul of F8_13 10.6 ul of F8_14 12.1 ul of F8_15 13.6 ul of F8_16 15.2 ul of F8_17 16.7 ul of F8_18	1.8 ul of F9_9 3.6 ul of F9_10 5.5 ul of F9_11 7.3 ul of F9_12 9.1 ul of F9_13 10.9 ul of F9_14 12.7 ul of F9_15 14.5 ul of F9_16 16.4 ul of F9_17 18.2 ul of F9_18	2.2 ul of F10_10 4.4 ul of F10_11 6.7 ul of F10_12 8.9 ul of F10_13 11.1 ul of F10_14 13.3 ul of F10_15 15.6 ul of F10_16 17.8 ul of F10_17 20.0 ul of F10_18
<b>Ligation Step #11</b>	<b>Ligation Step #12</b>	<b>Ligation Step #13</b>	<b>Ligation Step #14</b>	<b>Ligation Step #15</b>
2.8 ul of F11_11 5.6 ul of F11_12 8.3 ul of F11_13 11.1 ul of F11_14 13.9 ul of F11_15 16.7 ul of F11_16 19.4 ul of F11_17 22.2 ul of F11_18	3.6 ul of F12_12 7.1 ul of F12_13 10.7 ul of F12_14 14.3 ul of F12_15 17.9 ul of F12_16 21.4 ul of F12_17 25.0 ul of F12_18	4.8 ul of F13_13 9.5 ul of F13_14 14.3 ul of F13_15 19.0 ul of F13_16 23.8 ul of F13_17 28.6 ul of F13_18	6.7 ul of F14_14 13.3 ul of F14_15 20.0 ul of F14_16 26.7 ul of F14_17 33.3 ul of F14_18	10.0 ul of F15_15 20.0 ul of F15_16 30.0 ul of F15_17 40.0 ul of F15_18
<b>Ligation Step #16</b>	<b>Ligation Step #17</b>	<b>Ligation Step #18</b>		
16.7 ul of F16_16 33.3 ul of F16_17	33.3 ul of F17_17 66.7 ul of F17_18	100.0 ul of F18_18		

50.0 ul of F16_18				
-------------------	--	--	--	--

Carrying out the preceding ligation reactions results in a calculated PDF. Thus, the system can then adjust the volumes of each pre-ligated fragment during a further round of simulated reassembly until the PDF matches the desired probability function. The majority of progeny molecules only have one or two crossover events. Adjusting the quantities of the ligation reactions, as shown below will skew the PDF so that it moves towards progeny molecules having more crossover events.

One skilled in the art will readily appreciate that the present invention is well adapted to carry out the objects and obtain the ends and advantages mentioned as well as those inherent therein. The methods described herein are presently representative of exemplary aspects and are not intended as limitations on the scope of the invention. Changes therein and other uses will occur to those skilled in the art which are encompassed within the spirit of the invention and are defined by the scope of the claims.

## LITERATURE CITED

Unless otherwise indicated, all references cited herein (supra and infra) are incorporated by reference in their entirety.

- Altting-Mecs MA and Short JM: Polycos vectors: a system for packaging filamentous phage and phagemid vectors using lambda phage packaging extracts. *Gene* 137:1, 93-100, 1993.
- Arkin AP and Youvan DC: An algorithm for protein engineering: simulations of recursive ensemble mutagenesis. *Proc Natl Acad Sci USA* 89(16):7811-7815, (Aug 15) 1992.
- Arnold FH: Protein engineering for unusual environments. *Current Opinion in Biotechnology* 4(4):450-455, 1993.
- Arslan T, Abraham AT, Hecht SM: Structurally altered substrates for DNA topoisomerase I. Effects of inclusion of a single 3'-deoxynucleotide within the scissile strand. *Nucleosides Nucleotides* 1998 Jan-Mar;17(1-3):515-30.
- Aupeix K, Toulme JJ: Binding of chemically-modified oligonucleotides to the double-stranded stem of an RNA hairpin. *Nucleosides Nucleotides* 1999 Jun-Jul;18(6-7):1647-50.
- Ausubel FM, et al Editors. Current Protocols in Molecular Biology, Vols. 1 and 2 and supplements. (a.k.a. "The Red Book") Greene Publishing Assoc., Brooklyn, NY, ©1987.
- Ausubel FM, et al Editors. Current Protocols in Molecular Biology, Vols. 1 and 2 and supplements. (a.k.a. "The Red Book") Greene Publishing Assoc., Brooklyn, NY, ©1989.

- Ausubel FM, et al Editors. Short Protocols in Molecular Biology: A Compendium of Methods from Current Protocols in Molecular Biology. Greene Publishing Assoc., Brooklyn, NY, ©1989.
- Ausubel FM, et al Editors. Short Protocols in Molecular Biology: A Compendium of Methods from Current Protocols in Molecular Biology, 2<sup>nd</sup> Edition. Greene Publishing Assoc., Brooklyn, NY, ©1992.
- Barbas CF 3d, Bain JD, Hoekstra DM, Lerner RA: Semisynthetic combinatorial antibody libraries: a chemical solution to the diversity problem. *Proc Natl Acad Sci USA* 89(10):4457-4461, 1992.
- Bardwell AJ, Bardwell L, Johnson DK, Friedberg EC: Yeast DNA recombination and repair proteins Rad1 and Rad10 constitute a complex in vivo mediated by localized hydrophobic domains. *Mol Microbiol* 8(6):1177-1188, 1993.
- Barret AJ, et al., eds.: Enzyme Nomenclature: Recommendations of the Nomenclature Committee of the International Union of Biochemistry and Molecular Biology. San Diego: Academic Press, Inc., 1992.
- Bartel P, Chien CT, Sternglanz R, Fields S: Elimination of false positives that arise in using the two-hybrid system. *Biotechniques* 14(6):920-924, 1993.
- Bazzanini R, Manfredini S, Durini E, Groschel B, Cinatl J, Balzarini J, De Clercq E, Imbach JL, Perigaud C, Gosselin G: Prodrugs of Ara-CMP and Ara-AMP with a S-acyl-2-thioethyl (SATE) biolabile phosphate protecting group: synthesis and biological evaluation. *Nucleosides Nucleotides* 1999 Apr-May;18(4-5):971-2.
- Beaudry AA and Joyce GF: Directed evolution of an RNA enzyme. *Science* 257(5070):635-641, 1992.
- Berger and Kimmel, Methods in Enzymology, Volume 152, Guide to Molecular Cloning Techniques. Academic Press, Inc., San Diego, CA, ©1987. (Cumulative Subject Index: Volumes 135-139, 141-167, 1990, 272 pp.)
- Bevan M: Binary Agrobacterium vectors for plant transformation. *Nucleic Acids Research* 12(22):8711-21, 1984.
- Biocca S, Pierandrei-Amaldi P, Cattaneo A: Intracellular expression of anti-p21ras single chain Fv fragments inhibits meiotic maturation of xenopus oocytes. *Biochem Biophys Res Commun* 197(2):422-427, 1993.
- Bird et al. *Plant Mol Biol* 11:651, 1988..
- Blackburn GM, Liu X, Rosler A, Brenner C: Two hydrolase resistant analogues of diadenosine 5',5'''-P1,P3-triphosphate for studies with Fhit, the human fragile histidine triad protein. *Nucleosides Nucleotides* 1998 Jan-Mar;17(1-3):301-8.
- Bogerd HP, Fridell RA, Blair WS, Cullen BR: Genetic evidence that the Tat proteins of human immunodeficiency virus types 1 and 2 can multimerize in the eukaryotic cell nucleus. *J Virol* 67(8):5030-5034, 1993.
- Boyce COL, ed.: Novo's Handbook of Practical Biotechnology. 2<sup>nd</sup> ed. Bagsvaerd, Denmark, 1986.
- Brederode FT, Koper-Zawrtthoff EC, Bol JF: Complete nucleotide sequence of alfalfa mosaic virus RNA 4. *Nucleic Acids Research* 8(10):2213-23, 1980.
- Breitling F, Dubel S, Seehaus T, Klewinghaus I, Little M: A surface expression vector for antibody screening. *Gene* 104(2):147-153, 1991.

- Bridson PK, Lin X, Melman N, Ji XD, Jacobson KA: Synthesis and adenosine receptor affinity of 7-beta-D-ribofuranosylxanthine. *Nucleosides Nucleotides* 1998 Apr;17(4):759-68.
- Brodin P, Gottikh M, Auclair C, Mouscadet JF: Inhibition of HIV-1 integration by mono- & bi-functionalized triple helix forming oligonucleotides. *Nucleosides Nucleotides* 1999 Jun-Jul;18(6-7):1717-8.
- Brown NL, Smith M: Cleavage specificity of the restriction endonuclease isolated from *Haemophilus gallinarum* (Hga I). *Proc Natl Acad Sci U S A* 74(8):3213-6, (Aug) 1977.
- Burton DR, Barbas CF 3d, Persson MA, Koenig S, Chanock RM, Lerner RA: A large array of human monoclonal antibodies to type I human immunodeficiency virus from combinatorial libraries of asymptomatic seropositive individuals. *Proc Natl Acad Sci U S A* 88(22):10134-7, (Nov 15) 1991.
- Caldwell RC and Joyce GF: Randomization of genes by PCR mutagenesis. *PCR Methods Appl* 2(10):28-33, 1992.
- Caton AJ and Koprowski H: Influenza virus hemagglutinin-specific antibodies isolated from a combinatorial expression library are closely related to the immune response of the donor. *Proc Natl Acad Sci USA* 87(16):6450-6454, 1990.
- Chakraborty T, Martin JF, Olson EN: Analysis of the oligomerization of myogenin and E2A products in vivo using a two-hybrid assay system. *J Biol Chem* 267(25):17498-501, 1992.
- Chang CN, Landolfi NF, Queen C: Expression of antibody Fab domains on bacteriophage surfaces. Potential use for antibody selection. *J Immunol* 147(10):3610-4, (Nov 15) 1991.
- Chaudhary VK, Batra JK, Gallo MG, Willingham MC, FitzGerald DJ, Pastan I: A rapid method of cloning functional variable-region antibody genes in *Escherichia coli* as single-chain immunotoxins. *Proc Natl Acad Sci USA* 87(3):1066-1070, 1990.
- Chien CT, Bartel PL, Sternglanz R, Fields S: The two-hybrid system: a method to identify and clone genes for proteins that interact with a protein of interest. *Proc Natl Acad Sci USA* 88(21):9578-9582, 1991.
- Chiswell DJ, McCafferty J: Phage antibodies: will new 'coliclonal' antibodies replace monoclonal antibodies? *Trends Biotechnol* 10(3):80-84, 1992.
- Chothia C and Lesk AM: Canonical structures for the hypervariable regions of immunoglobulins. *J Mol Biol* 196(4):901-917, 1987.
- Chothia C, Lesk AM, Tramontano A, Levitt M, Smith-Gill SJ, Air G, Sheriff S, Padlan EA, Davies D, Tulip WR, et al: Conformations of immunoglobulin hypervariable regions. *Nature* 342(6252):877-883, 1989.
- Clackson T, Hoogenboom HR, Griffiths AD, Winter G: Making antibody fragments using phage display libraries. *Nature* 352(6336):624-628, 1991.
- Conrad M, Topal MD: DNA and spermidine provide a switch mechanism to regulate the activity of restriction enzyme *Nae I*. *Proc Natl Acad Sci U S A* 86(24):9707-11, (Dec) 1989.
- Coruzzi G, Broglie R, Edwards C, Chua NH: Tissue-specific and light-regulated expression of a pea nuclear gene encoding the small subunit of ribulose-1,5-bisphosphate carboxylase. *EMBO J* 3(8):1671-9, 1984.

- Creighton TE: *Proteins Structures and Molecular Principles*. New York: W. H. Freeman and Co., 1984.
- Dasmahapatra B, DiDomenico B, Dwyer S, Ma J, Sadowski I, Schwartz J: A genetic system for studying the activity of a proteolytic enzyme. *Proc Natl Acad Sci USA* 89(9):4159-4162, 1992.
- Davis LG, Dibner MD, Battey JF. Basic Methods in Molecular Biology. Elsevier, New York, NY, ©1986.
- De Clercq E: Carbocyclic adenosine analogues as S-adenosylhomocysteine hydrolase inhibitors and antiviral agents: recent advances. *Nucleosides Nucleotides* 1998 Jan-Mar;17(1-3):625-34.
- de Zwart M, Link R, von Frijtag Drabbe Kunzel JK, Cristalli G, Jacobson KA, Townsend-Nicholson A, IJzerman AP: A functional screening of adenosine analogues at the adenosine A2B receptor: a search for potent agonists. *Nucleosides Nucleotides* 1998 Jun;17(6):969-85.
- Delegrave S and Youvan DC. *Biotechnology Research* 11:1548-1552, 1993.
- DeLong EF, Wu KY, Prezelin BB, Jovine RV: High abundance of Archaea in Antarctic marine picoplankton. *Nature* 371(6499):695-697, 1994.
- Deng SJ, MacKenzie CR, Sadowska J, Michniewicz J, Young NM, Bundle Dr, Narang SA: Selection of antibody single-chain variable fragments with improved carbohydrate binding by phage display. *J Biol Chem* 269(13):9533-9538, 1994.
- Drauz K, Waldman H, eds.: Enzyme Catalysis in Organic Synthesis: A Comprehensive Handbook. Vol. 1. New York: VCH Publishers, 1995.
- Drauz K, Waldman H, eds.: Enzyme Catalysis in Organic Synthesis: A Comprehensive Handbook. Vol. 2. New York: VCH Publishers, 1995.
- Duan L, Bagasra O, Laughlin MA, Oakes JW, Pomerantz RJ: Potent inhibition of human immunodeficiency virus type 1 replication by an intracellular anti-Rev single-chain antibody. *Proc Natl Acad Sci USA* 91(11):5075-5079, 1994.
- Durfee T, Becherer K, Chen PL, Yeh SH, Yang Y, Kilburn AE, Lee WH, Elledge SJ: The retinoblastoma protein associates with the protein phosphatase type 1 catalytic subunit. *Genes Dev* 7(4):555-569, 1993.
- Egron D, Arzumanov AA, Dyatkina NB, Krayevsky A, Imbach JL, Aubertin AM, Gosselin G, Perigaud C: Synthesis, anti-HIV activity and stability studies of 3'-azido-2',3'-dideoxythymidine 5'-fluorophosphate. *Nucleosides Nucleotides* 1999 Apr-May;18(4-5):983-4
- Ellington AD and Szostak JW: In vitro selection of RNA molecules that bind specific ligands. *Nature* 346(6287):818-822, 1990.
- Fields S and Song O: A novel genetic system to detect protein-protein interactions. *Nature* 340(6230):245-246, 1989.
- Firek S, Draper J, Owen MR, Gandeche A, Cockburn B, Whitlam GC: Secretion of a functional single-chain Fv protein in transgenic tobacco plants and cell suspension cultures. *Plant Mol Biol* 23(4):861-870, 1993.
- Forsblom S, Rigler R, Ehrenberg M, Philipson L: Kinetic studies on the cleavage of adenovirus DNA by restriction endonuclease Eco RI. *Nucleic Acids Res* 3(12):3255-69, (Dec) 1976.

- Foster GD, Taylor SC, eds.: Plant Virology Protocols: From Virus Isolation to Transgenic Resistance. Methods in Molecular Biology, Vol. 81. New Jersey: Humana Press Inc., 1998.
- Franks F, ed.: Protein Biotechnology: Isolation, Characterization, and Stabilization. New Jersey: Humana Press Inc., 1993.
- Germino FJ, Wang ZX, Weissman SM: Screening for in vivo protein-protein interactions. - *Proc Natl Acad Sci USA* 90(3):933-937, 1993.
- Gianolio DA, McLaughlin LW: Synthesis and triplex forming properties of pyrimidine derivative containing extended functionality. *Nucleosides Nucleotides* 1999 Aug;18(8):1751-69.
- Gingeras TR, Brooks JE: Cloned restriction/modification system from *Pseudomonas aeruginosa*. *Proc Natl Acad Sci USA* 80(2):402-6, 1983 (Jan).
- Gluzman Y: SV40-transformed simian cells support the replication of early SV40 mutants. *Cell* 23(1):175-182, 1981.
- Godfrey T, West S, eds.: Industrial Enzymology. 2<sup>nd</sup> ed. London: Macmillan Press Ltd, 1996.
- Gottikh MB, Volkov EM, Romanova EA, Oretskaya TS, Shabarova ZA: Synthesis of oligonucleotide-intercalator conjugates capable to inhibit HIV-1 DNA integration. *Nucleosides Nucleotides* 1999 Jun-Jul;18(6-7):1645-6.
- Gottschalk G: Bacterial Metabolism. 2<sup>nd</sup> ed. New York: Springer-Verlag Inc., 1986.
- Gresshoff PM, ed.: Technology Transfer of Plant Biotechnology. Current Topics in Plant Molecular Biology. Boca Raton: CRC Press, 1997.
- Griffin HG, Griffin AM, eds.: PCR Technology: Current Innovations. Boca Raton: CRC Press, Inc., 1994.
- Gruber M, Schodin BA, Wilson ER, Kranz DM: Efficient tumor cell lysis mediated by a bispecific single chain antibody expressed in *Escherichia coli*. *J Immunol* 152(11):5368-5374, 1994.
- Guarente L: Strategies for the identification of interacting proteins. *Proc Natl Acad Sci USA* 90(5):1639-1641, 1993.
- Guilley H, Dudley RK, Jonard G, Balazs E, Richards KE: Transcription of Cauliflower mosaic virus DNA: detection of promoter sequences, and characterization of transcripts. *Cell* 30(3):763-73, 1982.
- Hansen G, Chilton MD: Lessons in gene transfer to plants by a gifted microbe. *Curr Top Microbiol Immunol* 240:21-57, 1999.
- Hardy CF, Sussel L, Shore D: A RAP1-interacting protein involved in transcriptional silencing and telomere length regulation. *Genes Dev* 6(5):801-814, 1992.
- Hartmann HT, et al.: Plant Propagation: Principles and Practices. 6<sup>th</sup> ed. New Jersey: Prentice Hall, Inc., 1997.
- Hawkins RE and Winter G: Cell selection strategies for making antibodies from variable gene libraries: trapping the memory pool. *Eur J Immunol* 22(3):867-870, 1992.
- Holvoet P, Laroche Y, Lijnen HR, Van Hoef B, Brouwers E, De Cock F, Lauwereys M, Gansemans Y, Collen D: Biochemical characterization of single-chain chimeric plasminogen activators consisting of a single-chain Fv fragment of a fibrin-specific antibody and single-chain urokinase. *Eur J Biochem* 210(3):945-952, 1992.



- Honjo T, Alt FW, Rabbitts TH (eds): *Immunoglobulin genes*. Academic Press: San Diego, CA, pp. 361-368, ©1989.
- Hoogenboom HR, Griffiths AD, Johnson KS, Chiswell DJ, Judson P, Winter G: Multi-subunit proteins on the surface of filamentous phage: methodologies for displaying antibody (Fab) heavy and light chains. *Nucleic Acids Res* 19(15):4133-4137, 1991.
- Hotoda H, Koizumi M, Ohmine T, Furukawa H, Nishigaki T, Abe K, Kosaka T, Tsutsumi S, Sone J, Kaneko M: Biologically active oligodeoxyribonucleotides. 10: anti-HIV-1 activity and stability of modified hexanucleotides containing glycerol-skeleton. *Nucleosides Nucleotides* 1998 Jan-Mar;17(1-3):243-52.
- Huse WD, Sastry L, Iverson SA, Kang AS, Alting-Mees M, Burton DR, Benkovic SJ, Lerner RA: Generation of a large combinatorial library of the immunoglobulin repertoire in phage lambda. *Science* 246(4935):1275-1281, 1989.
- Huston JS, Levinson D, Mudgett-Hunter M, Tai MS, Novotney J, Margolies MN, Ridge RJ, Brucoleri RE, Haber E, Crea R, et al: Protein engineering of antibody binding sites: recovery of specific activity in an anti-digoxin single-chain Fv analogue produced in *Escherichia coli*. *Proc Natl Acad Sci USA* 85(16):5879-5883, 1988.
- Ivan Lefkovits, Editor. *Immunology methods manual : the comprehensive sourcebook of techniques*. Academic Press, San Diego, ©1997.
- Iwabuchi K, Li B, Bartel P, Fields S: Use of the two-hybrid system to identify the domain of p53 involved in oligomerization. *Oncogene* 8(6):1693-1696, 1993.
- Jackson AL, Pahl PM, Harrison K, Rosamond J, Sclafani RA: Cell cycle regulation of the yeast Cdc7 protein kinase by association with the Dbf4 protein. *Mol Cell Biol* 13(5):2899-2908, 1993.
- Johnson S and Bird RE: *Methods Enzymol* 203:88, 1991.
- JP10113194; Filed 19971022, Published 19980506. Donnelly, JJ ;Dwarki, VJ ;Liu, MA ;Montgomery, DL ;Parker, S ;Shiver, JW ;Ulmer JB: Nucleic Acid Preparation.
- Kabat et al: Sequences of Proteins of Immunological Interest, 4th Ed. U.S. Department of Health and Human Services, Bethesda, MD (1987)
- Kang AS, Barbas CF, Janda KD, Benkovic SJ, Lerner RA: Linkage of recognition and replication functions by assembling combinatorial antibody Fab libraries along phage surfaces. *Proc Natl Acad Sci USA* 88(10):4363-4366, 1991.
- Kang SH, Sinhababu AK, Cho MJ: Synthesis and biological activity of bis(pivaloyloxymethyl) ester of 2'-azido-2'-deoxyuridine 5'-monophosphate. *Nucleosides Nucleotides* 1998 Jun;17(6):1089-98.
- Kettleborough CA, Ansell KH, Allen RW, Rosell-Vives E, Gussow DH, Bendig MM: Isolation of tumor cell-specific single-chain Fv from immunized mice using phage-antibody libraries and the re-construction of whole antibodies from these antibody fragments. *Eur J Immunol* 24(4):952-958, 1994.

- Krayevsky A, Arzumanov A, Shirokova E, Dyatkina N, Victorova L, Jasko M, Alexandrova L: dNTP modified at triphosphate residues: substrate properties towards DNA polymerases and stability in human serum. *Nucleosides Nucleotides* 1998 Jan-Mar;17(1-3):681-93.
- Krayevsky AA, Dyatkina NB, Semizarov DG, Victorova LS, Shirokova EA, Theil F, Von Janta Lipinski MJ, Gosselin G, Imbach JL: Reasons and limits of substrate activity of modified L-dNTP in DNA biosynthesis. *Nucleosides Nucleotides* 1999 Apr-May;18(4-5):863-4.
- Kruger DH, Barcak GJ, Reuter M, Smith HO: EcoRII can be activated to cleave refractory DNA recognition sites. *Nucleic Acids Res* 16(9):3997-4008, (May 11) 1988.
- Kvasyuk EI, Mikhailopulo IA, Suhadolnik RJ, Henderson EE, Muto NF, Iacono KT, Homon J, Pfeleiderer W: Synthesis and biological activity of 2',5'-oligoadenylate trimers containing 5'-terminal 5'-amino-5'-deoxy- and 5'-amino-3',5'-dideoxyadenosine derivatives. *Nucleosides Nucleotides* 1999 Jun-Jul;18(6-7):1483-4.
- Lalo D, Carles C, Sentenac A, Thuriaux P: Interactions between three common subunits of yeast RNA polymerases I and III. *Proc Natl Acad Sci USA* 90(12):5524-5528, 1993.
- Laskowski M Sr: Purification and properties of venom phosphodiesterase. *Methods Enzymol* 65(1):276-84, 1980.
- Lefkovits I and Pernis B, Editors. *Immunological Methods*, Vols. I and II. Academic Press, New York, NY. Also Vol. III published in Orlando and Vol. IV published in San Diego. ©1979-.
- Lerner RA, Kang AS, Bain JD, Burton DR, Barbas CF 3d: Antibodies without immunization. *Science* 258(5086):1313-1314, 1992.
- Leung, D.W., et al, *Technique*, 1:11-15, 1989.
- Li B and Fields S: Identification of mutations in p53 that affect its binding to SV40 large T antigen by using the yeast two-hybrid system. *FASEB J* 7(10):957-963, 1993.
- Lilley GG, Doelzal O, Hillyard CJ, Bernard C, Hudson PJ: Recombinant single-chain antibody peptide conjugates expressed in *Escherichia coli* for the rapid diagnosis of HIV. *J Immunol Methods* 171(2):211-226, 1994.
- Liu J, Skradis A, Kolar C, Kolath J, Anderson J, Lawson T, Talmadge J, Gmeiner WH: Increased cytotoxicity and decreased in vivo toxicity of FdUMP[10] relative to 5-FU. *Nucleosides Nucleotides* 1999 Aug;18(8):1789-802.
- Lowman HB, Bass SH, Simpson N, Wells JA: Selecting high-affinity binding proteins by monovalent phage display. *Biochemistry* 30(45):10832-10838, 1991.
- Luban J, Bossolt KL, Franke EK, Kalpana GV, Goff SP: Human immunodeficiency virus type 1 Gag protein binds to cyclophilins A and B. *Cell* 73(6):1067-1078, 1993.
- Lutz MJ, Will DW, Breipohl G, Benner SA, Uhlmann E: Synthesis of a monocharged peptide nucleic acid (PNA) analog and its recognition as substrate by DNA polymerases. *Nucleosides Nucleotides* 1999 Mar;18(3):393-401.
- Madura K, Dohmen RJ, Varshavsky A: N-recognition/Ubc2 interactions in the N-end rule pathway. *J Biol Chem* 268(16):12046-54, (Jun 5) 1993.
- Marks JD, Griffiths Ad, Malmqvist M, Clackson TP, Bye JM, Winter G: By-passing immunization: building high affinity human antibodies by chain shuffling. *Biotechnology (N Y)* 10(7):779-783, 1992.

- Marks JD, Hoogenboom HR, Bonnert TP, McCafferty J, Griffiths AD, Winter G: Bypassing immunization. Human antibodies from V-gene libraries displayed on phage. *J Mol Biol* 222(3):581-597, 1991.
- Marks JD, Hoogenboom HR, Griffiths AD, Winter G: Molecular evolution of proteins on filamentous phage. Mimicking the strategy of the immune system. *J Biol Chem* 267(23):16007-16010, 1992.
- Maxam AM, Gilbert W: Sequencing end-labeled DNA with base-specific chemical cleavages. *Methods Enzymol* 65(1):499-560, 1980.
- McCafferty J, Griffiths AD, Winter G, Chiswell DJ: Phage antibodies: filamentous phage displaying antibody variable domains. *Nature* 348(6301):552-554, 1990.
- Miller JH. A Short Course in Bacterial Genetics: A Laboratory Manual and Handbook for Escherichia coli and Related Bacteria (see inclusively p. 445). Cold Spring Harbor Laboratory Press, Plainview, NY, ©1992.
- Milne GT and Weaver DT: Dominant negative alleles of RAD52 reveal a DNA repair/recombination complex including Rad51 and Rad52. *Genes Dev* 7(9):1755-1765, 1993.
- Monaco V, van de Wetering KI, Meeuwenoord NJ, van den Elst HA, Stuivenberg HR, Visse R, van der Kaaden JC, Moolenaar GF, Verhoeven EE, Goosen N, van der Marel GA, van Boom JH: Synthesis and biological evaluation of modified DNA fragments for the study of nucleotide excision repair in E. coli. *Nucleosides Nucleotides* 1999 Jun-Jul;18(6-7):1339-41.
- Morozova OV, Kolpashchikov DM, Ivanova TM, Godovikova TS: Synthesis of new photocross-linking 5-C-base-substituted UTP analogs and their application in highly selective affinity labelling of the tick-borne encephalitis virus RNA replicase proteins. *Nucleosides Nucleotides* 1999 Jun-Jul;18(6-7):1513-4.
- Mullinax RL, Gross EA, Amberg JR, Hay BN, Hogrefe HH, Kubit MM, Greener A, Alting-Mees M, Ardourel D, Short JM, et al: Identification of human antibody fragment clones specific for tetanus toxoid in a bacteriophage lambda immunoexpression library. *Proc natl Acad Sci USA* 87(20):8095-9099, 1990.
- Nath K, Azzolina BA: in *Gene Amplification and Analysis* (ed. Chirikjian JG), vol. 1, p. 113, Elsevier North Holland, Inc., New York, New York, ©1981.
- Needleman SB and Wunsch CD: A general method applicable to the search for similarities in the amino acid sequence of two proteins. *J Mol Biol* 48(3):443-453, 1970.
- Nelson M, Christ C, Schildkraut I: Alteration of apparent restriction endonuclease recognition specificities by DNA methylases. *Nucleic Acids Res* 12(13):5165-73, 1984 (Jul 11).
- Nguyen-Ba N, Chan L, Quimpere M, Turcotte N, Lee N, Mitchell H, Bedard J: Design and SAR study of a novel class of nucleotide analogues as potent anti-HCMV agents. *Nucleosides Nucleotides* 1999 Apr-May;18(4-5):821-7.
- Nicholls PJ, Johnson VG, Andrew SM, Hoogenboom HR, Raus JC, Youle RJ: Characterization of single-chain antibody (sFv)-toxin fusion proteins produced in vitro in rabbit reticulocyte lysate. *J Biol Chem* 268(7):5302-5308, 1993.
- Oller AR, Vanden Broek W, Conrad M, Topal MD: Ability of DNA and spermidine to affect the activity of restriction endonucleases from several bacterial species. *Biochemistry* 30(9):2543-9, (Mar 5) 1991.

- Owen MRL, Pen J: Transgenic Plants: A Production System for Industrial and Pharmaceutical Proteins. Chichester: John Wiley & Sons, 1996.
- Owens RJ and Young RJ: The genetic engineering of monoclonal antibodies. *J Immunol Methods* 168(2):149-165, 1994.
- Pandolfi D, Rauzi F, Capobianco ML: Evaluation of different types of end-capping modifications on the stability of oligonucleotides toward 3'- and 5'-exonucleases. *Nucleosides Nucleotides* 1999 Sep;18(9):2051-69.
- Pankiewicz KW, Lesiak-Watanabe K: Novel mycophenolic adenine bis(phosphonate)s as potent anticancer agents and inducers of cells differentiation. *Nucleosides Nucleotides* 1999 Apr-May;18(4-5):927-32.
- Pearson WR and Lipman DJ: Improved tools for biological sequence comparison. *Proc Natl Acad Sci USA* 85(8):2444-2448, 1988.
- Pein CD, Reuter M, Meisel A, Cech D, Kruger DH: Activation of restriction endonuclease EcoRII does not depend on the cleavage of stimulator DNA. *Nucleic Acids Res* 19(19):5139-42, (Oct 11) 1991.
- Perrin DM, Garestier T, Helene C: Expanding the catalytic repertoire of nucleic acid catalysts: simultaneous incorporation of two modified deoxyribonucleoside triphosphates bearing ammonium and imidazolyl functionalities. *Nucleosides Nucleotides* 1999 Mar;18(3):377-91.
- Persson MA, Caothien RH, Burton DR: Generation of diverse high-affinity human monoclonal antibodies by repertoire cloning. *Proc Natl Acad Sci USA* 88(6):2432-2436, 1991.
- Perun TJ, Propst CL, eds.: Computer-Aided Drug Design: Methods and Applications. New York: Marcel Dekker, Inc., 1989.
- Pfundheller HM, Koshkin AA, Olsen CE, Wengel J: Evaluation of oligonucleotides containing two novel 2'-O-methyl modified nucleotide monomers: a 3'-C-allyl and a 2'-O,3'-C-linked bicyclic derivative. *Nucleosides Nucleotides* 1999 Sep;18(9):2017-30.
- Qiang BQ, McClelland M, Poddar S, Spokauskas A, Nelson M: The apparent specificity of NotI (5'-GCGGCCGC-3') is enhanced by M.FnuDII or M.BepI methyltransferases (5'-mCGCG-3'): cutting bacterial chromosomes into a few large pieces. *Gene* 88(1):101-5, (Mar 30) 1990.
- Queen C, Foster J, Stauber C, Stafford J: Cell-type specific regulation of a kappa immunoglobulin gene by promoter and enhance elements. *Immunol Rev* 89:49-68, 1986.
- Raleigh EA, Wilson G: Escherichia coli K-12 restricts DNA containing 5-methylcytosine. *Proc Natl Acad Sci U S A* 83(23):9070-4, (Dec) 1986.
- Ramasamy KS, Stoisavljevic V: Synthesis and biophysical studies of modified oligonucleotides containing acyclic amino alcohol nucleoside analogs. *Nucleosides Nucleotides* 1999 Aug;18(8):1845-61.
- Reidhaar-Olson JF and Sauer RT: Combinatorial cassette mutagenesis as a probe of the informational content of protein sequences. *Science* 241(4861):53-57, 1988.
- Riechmann L and Weill M: Phage display and selection of a site-directed randomized single-chain antibody Fv fragment for its affinity improvement. *Biochemistry* 32(34):8848-8855, 1993.

- Roberts RJ, Macelis D: REBASE--restriction enzymes and methylases. *Nucleic Acids Res* 24(1):223-35, (Jan 1) 1996.
- Ryan AJ, Royal CL, Hutchinson J, Shaw CH: Genomic sequence of a 12S seed storage protein from oilseed rape (*Brassica napus* c.v. jet neuf). *Nucl Acids Res* 17(9):3584, 1989.
- Sambrook J, Fritsch EF, Maniatis T. Molecular Cloning: A Laboratory Manual. Cold Spring Harbor Laboratory Press, Cold Spring Harbor, NY, ©1982.
- Sambrook J, Fritsch EF, Maniatis T. Molecular Cloning: A Laboratory Manual. Second Edition. Cold Spring Harbor Laboratory Press, Cold Spring Harbor, NY, ©1989.
- Schinazi RF, Lesnikowski ZJ: Boron containing oligonucleotides. *Nucleosides Nucleotides* 1998 Jan-Mar;17(1-3):635-47.
- Scopes RK. Protein Purification: Principles and Practice. Springer-Verlag, New York, NY, © 1982.
- Secrist JA 3rd, Parker WB, Allan PW, Bennett LL Jr, Waud WR, Truss JW, Fowler AT, Montgomery JA, Ealick SE, Wells AH, Gillespie GY, Gadi VK, Sorscher EJ: Gene therapy of cancer: activation of nucleoside prodrugs with *E. coli* purine nucleoside phosphorylase. *Nucleosides Nucleotides* 1999 Apr-May;18(4-5):745-57.
- Segel IH: Enzyme Kinetics: Behavior and Analysis of Rapid Equilibrium and Steady-State Enzyme Systems. New York: John Wiley & Sons, Inc., 1993.
- Shirokova EA, Shipitsin AV, Victorova LS, Dyatkina NB, Goryunova LE, Beabealashvili RS, Hamilton CJ, Roberts SM, Krayevsky AA: Modified nucleoside 5'-triphosphonates as a new type of antiviral agents. *Nucleosides Nucleotides* 1999 Apr-May;18(4-5):1027-8.
- Silver SC and Hunt SW 3d: Techniques for cloning cDNAs encoding interactive transcriptional regulatory proteins. *Mol Biol Rep* 17(3):155-165, 1993.
- Smith TF, Waterman MS, Fitch WM: Comparative biosequence metrics. *J Mol Evol* S18(1):38-46, 1981.
- Smith TF, Waterman MS. *Adv Appl Math* 2: 482-end of article, 1981.
- Smith TF, Waterman MS: Identification of common molecular subsequences. *J Mol Biol* 147(1):195-7, (Mar 25) 1981.
- Smith TF, Waterman MS: Overlapping genes and information theory. *J Theor Biol* 91(2):379-80, (Jul 21) 1981.
- Srivastava TK, Friedhoff P, Pingoud A, Katti SB: Application of oligonucleoside methylphosphonates in the studies on phosphodiester hydrolysis by *Serratia* endonuclease. *Nucleosides Nucleotides* 1999 Sep;18(9):1945-60.
- Stattel JM, Yanachkov I, Wright GE: Synthesis and biochemical study of N2-(p-n-butylphenyl)-2'-deoxyguanosine 5'-(alpha,beta-imido)triphosphate (BuPdGMPNHPP): a non-substrate inhibitor of B family DNA polymerases. *Nucleosides Nucleotides* 1998 Aug;17(8):1505-13.
- Staudinger J, Perry M, Elledge SJ, Olson EN: Interactions among vertebrate helix-loop-helix proteins in yeast using the two-hybrid system. *J Biol Chem* 268(7):4608-4611, 1993.

- Stemmer WP, Morris SK, Wilson BS: Selection of an active single chain Fv antibody from a protein linker library prepared by enzymatic inverse PCR. *Biotechniques* 14(2):256-265, 1993.
- Stemmer WP: DNA shuffling by random fragmentation and reassembly: in vitro recombination for molecular evolution. *Proc Natl Acad Sci USA* 91(22):10747-10751, 1994.
- Sun D, Hurley LH: Effect of the (+)-CC-1065-(N3-adenine)DNA adduct on in vitro DNA synthesis mediated by Escherichia coli DNA polymerase. *Biochemistry* 31:10, 2822-9, (Mar 17) 1992.
- Tague BW, Dickinson CD, Chrispeels MJ: A short domain of the plant vacuolar protein phytohemagglutinin targets invertase to the yeast vacuole. *Plant Cell* 2(6):533-46, (June) 1990.
- Takahashi N, Kobayashi I: Evidence for the double-strand break repair model of bacteriophage lambda recombination. *Proc Natl Acad Sci U S A* 87(7):2790-4, (Apr) 1990.
- Terato H, Morita H, Ohyama Y, Ide H: Novel modification of 5-formyluracil by cysteine derivatives in aqueous solution. *Nucleosides Nucleotides* 1998 Jan-Mar;17(1-3):131-41.
- Thiesen HJ and Bach C: Target Detection Assay (TDA): a versatile procedure to determine DNA binding sites as demonstrated on SP1 protein. *Nucleic Acids Res* 18(11):3203-3209, 1990.
- Thomas M, Davis RW: Studies on the cleavage of bacteriophage lambda DNA with EcoRI Restriction endonuclease. *J Mol Biol* 91(3):315-28, (Jan 25) 1975.
- Tingey SV, Walker EL, Coruzzi GM: Glutamine synthetase genes of pea encode distinct polypeptides which are differentially expressed in leaves, roots and nodules. *EMBO J* 6(1):1-9, 1987.
- Tomikawa A, Seno M, Sato-Kiyotaki K, Ohtsuki C, Hirai T, Yamaguchi T, Kawaguchi T, Yoshida S, Saneyoshi M: Synthetic nucleosides and nucleotides. 40. Selective inhibition of eukaryotic DNA polymerase alpha by 9-(beta-D-arabinofuranosyl)-2-(p-n-butylanilino) adenine 5'-triphosphate (BuAaraATP) and its 2'-up azido analog: synthesis and enzymatic evaluations. *Nucleosides Nucleotides* 1998 Jan-Mar;17(1-3):487-501.
- Topal MD, Thresher RJ, Conrad M, Griffith J: NaeI endonuclease binding to pBR322 DNA induces looping. *Biochemistry* 30(7):2006-10, (Feb. 19) 1991.
- Tramontano A, Chothia C, Lesk AM: Framework residue 71 is a major determinant of the position and conformation of the second hypervariable region in the VH domains of immunoglobulins. *J Mol Biol* 215(1):175-182, 1990.
- Tuerk C and Gold L: Systematic evolution of ligands by exponential enrichment: RNA ligands to bacteriophage T4 DNA polymerase. *Science* 249(4968):505-510, 1990.
- US5580859; Filed 19940318, Issued 19961203. Felgner, PL. ;Wolff, JA. ;Rhodes, GH. ;Malone, RW. ;Carson, DA.: Delivery of exogenous DNA sequences in a mammal.
- US5589466; Filed 19950126, Issued 19961231. Felgner, PL. ;Wolff, JA. ;Rhodes, GH. ;Malone, RW. ;Carson, DA.: Induction of a protective immune response in a mammal by injecting a DNA sequence.
- US5641665; Filed 19941128, Issued 19970624. Hobart, PM. ;Margalith, M ;Parker, SE. ;Khatibi, S: Plasmids suitable for IL-2 expression.

- US5693622; Filed 19950607, Issued 19971202. Wolff, JA. ;Duke, DJ. ;Felgner, PL.: Expression of exogenous polynucleotide sequences cardiac muscle of a mammal.
- US5703055; Filed 19940126, Issued 19971230. Felgner, PL. ;Wolff, JA ;Rhodes, GH. ;Malone, RW ;Carson, DA.: Generation of antibodies through lipid mediated DNA delivery.
- US5846946; Filed 19960614, Issued 19981208. Huebner, RC. ;Norman, JA. ;Liang, X ;Carner, KR. ;Barbour, AG. ;Luke, CJ.: Compositions and methods for administering Borrelia DNA.
- US5910488; Filed 19951201, Issued 19990608. Nabel, GJ. ;Nabel, EG. ;Lew, D ;Marquet, M: Plasmids suitable for gene therapy.
- USPN 4,683,195; Filed Feb. 7, 1986, Issued Jul 28, 1987. Mullis KB, Erlich HA, Arnheim N, Horn GT, Saiki RK, Scharf SJ: Process for Amplifying, Detecting, and/or Cloning Nucleic Acid Sequences.
- USPN 4,683,202; Filed Oct. 25, 1985, Issued Jul. 28, 1987. Mullis KB: Process for Amplifying Nucleic Acid Sequences.
- USPN 4,704,362; Filed Nov. 5, 1979, Issued Nov. 3, 1987. Itakura K, Riggs AD: Recombinant Cloning Vehicle Microbial Polypeptide Expression.
- van de Poll ML, Lafleur MV, van Gog F, Vrieling H, Meerman JH: N-acetylated and deacetylated 4'-fluoro-4-aminobiphenyl and 4-aminobiphenyl adducts differ in their ability to inhibit DNA replication of single-stranded M13 in vitro and of single-stranded phi X174 in Escherichia coli. *Carcinogenesis* 13(5):751-8, (May) 1992.
- Victorova LS, Semizarov DG, Shirokova EA, Alexandrova LA, Arzumanov AA, Jasko MV, Krayevsky AA: Human DNA polymerases and retroviral reverse transcriptases: selectivity in respect to dNTPs modified at triphosphate residues. *Nucleosides Nucleotides* 1999 Apr-May;18(4-5):1031-2.
- Vojtek AB, Hollenberg SM, Cooper JA: Mammalian Ras interacts directly with the serine/threonine kinase Raf. *Cell* 74(1):205-214, 1993.
- von Janta-Lipinski M, Gaertner K, Lehmann C, Scheer H, Schildt J, Matthes E: Protein and RNA of human telomerase as targets for modified oligonucleotides. *Nucleosides Nucleotides* 1999 Jun-Jul;18(6-7):1719-20
- Wenzler H, Mignery G, Fisher L, Park W: Sucrose-regulated expression of a chimeric potato tuber gene in leaves of transgenic tobacco plants. *Plant Mol Biol* 13(4):347-54, 1989.
- White JS, White DC: Source Book of Enzymes. Boca Raton: CRC Press, 1997.
- Williams and Barclay, in Immunoglobulin Genes, The Immunoglobulin Gene Superfamily
- Winnacker EL. From Genes to Clones: Introduction to Gene Technology. VCH Publishers, New York, NY, ©1987.
- Winter G and Milstein C: Man-made antibodies. *Nature* 349(6307):293-299, 1991.
- WO 88/08453; Filed Apr. 14, 1988, Published Nov. 3, 1988. Alakhov JB, Baranov, VI, Ovodov SJ, Ryabova LA, Spirin AS: Method of Obtaining Polypeptides in Cell-Free Translation System.
- WO 90/05785; Filed Nov. 15, 1989, Published May 31, 1990. Schultz P: Method for Site-Specifically Incorporating Unnatural Amino Acids into Proteins.

- WO 90/07003; Filed Jan. 27, 1989, Published June 28, 1990. Baranov VI, Morozov II, Spirin AS: Method for Preparative Expression of Genes in a Cell-free System of Conjugated Transcription/translation.
- WO 91/02076; Filed June 14, 1990, Published Feb. 21, 1991. Baranov VI, Ryabova LA, Yarchuk OB, Spirin AS: Method for Obtaining Polypeptides in a Cell-free System.
- WO 91/05058; Filed Oct. 5, 1989, Published Apr. 18, 1991. Kawasaki G: Cell-free Synthesis and Isolation of Novel Genes and Polypeptides.
- WO 91/17271; Filed May 1, 1990, Published Nov. 14, 1991. Dower WJ, Cwirla SE: Recombinant Library Screening Methods.
- WO 91/18980; Filed May 13, 1991, Published Dec. 12, 1991. Devlin JJ: Compositions and Methods for Identifying Biologically Active Molecules.
- WO 91/19818; Filed June 20, 1990, Published Dec. 26, 1991. Dower WJ, Cwirla SE, Barrett RW: Peptide Library and Screening Systems.
- WO 92/02536; Filed Aug. 1, 1991, Published Feb. 20, 1992. Gold L, Tuerk C: Systematic Polypeptide Evolution by Reverse Translation.
- WO 92/03918; Filed Aug. 28, 1991, Published Mar. 19, 1992. Lonberg N, Kay RM: Transgenic Non-human Animals Capable of Producing Heterologous Antibodies.
- WO 92/03918; Filed Aug. 28, 1991, Published Mar. 19, 1992. Lonberg N, Kay RM: Transgenic Non-human Animals Capable of Producing Heterologous Antibodies.
- WO 92/05258; Filed Sept. 17, 1991, Published Apr. 2, 1992. Fincher GB: Gene Encoding Barley Enzyme.
- WO 92/14843; Filed Feb. 21, 1992, Published Sept. 3, 1992. Toole JJ, Griffin LC, Bock LC, Latham JA, Muenchau DD, Krawczyk S: Aptamers Specific for Biomolecules and Method of Making.
- WO 93/08278; Filed Oct. 15, 1992, Published Apr. 29, 1993. Schatz PJ, Cull MG, Miller JF, Stemmer WP: Peptide Library and Screening Method.
- WO 93/12227; Filed Dec 17, 1992, Published June 24, 1993. Lonberg, N; Kay RM: Transgenic Non-human Animals Capable of Producing Heterologous Antibodies.
- WO 93/12227; Filed Dec. 17, 1992, Published June 24, 1993. Lonberg N, Kay RM: Transgenic Non-human Animals Capable of Producing Heterologous Antibodies.
- WO 94/25585; Filed Apr. 25, 1994, Published Nov. 10, 1994. Lonberg, N, Kay RM: Transgenic Non-human Animals Capable of Producing Heterologous Antibodies.
- WO 94/25585; Filed Apr. 25, 1994, Published Nov. 10, 1994. Lonberg N, Kay RM: Transgenic Non-human Animals Capable of Producing Heterologous Antibodies.
- WO 97/35966; Filed Mar. 20, 1997, Published Oct. 2, 1997. Minshull J, Stemmer WP: Methods and Compositions for Cellular and Metabolic Engineering.
- WO 98/31837; Filed Jan. 16, 1998, Published July 23, 1998. Delcardayre SB, Tobin MB, Stemmer WP, Minshull, J: Evolution of Whole Cells and Organisms by Recursive Sequence Recombination.
- WO 98/37223; Filed Feb. 18 1998, Published Aug. 27, 1998. Pang SZ, Gonsalves D, Jan FJ: DNA Construct to Confer Multiple Traits on Plants.
- WO9011092; Filed 19900321, A1 Published 19901004. Felgner, PL. ;Wolff, JA;Rhodes, GH. ;Malone, RW ;Carson, DA.: Expression Of Exogenous Polynucleotide Sequences In A Vertebrate.



- WO9314778; Filed 19930121, A1 Published 19930805. Rhodes, GH.; Dwarki, VJ.; Felgner, PL; Wang-Felgner, J; Manthorpe, M: Ex Vivo Gene Transfer.
- WO9421797; Filed 19940314, A1 Published 19940929. Donnelly, JJ.; Dwarki, VJ.; Liu, MA.; Montgomery, DL.; Parker, SE.; Shiver, JW.; Ulmer, JB.: Nucleic Acid Pharmaceuticals.
- WO9633736; Filed 19960426, A1 Published 19961031. Baruch DI; Pasloske BL; Howard, RJ: Malaria Peptides and Vaccines.
- WO9735992; Filed 19970317, A1 Published 19971002. Hobart, PM.; Liang, X: Tetracycline Inducible/Repressible Systems.
- WO9926663; Filed 19981120, A2 Published 19990603. Horton, H.; Parker, S; Manthorpe, M; Felgner, P: Treatment Of Cancer Using Cytokine-Expressing Polynucleotides And Compositions Therefor.
- WO9941368; Filed 19990210, A2 Published 19990819. Punnonen J, Stemmer WP, Whalen RG; Howard, R: Optimization of Immunomodulatory Properties of Genetic Vaccines.
- WO9941369; Filed 19990210, A2 Published 19990819. Punnonen J, Stemmer WP, Whalen RG; Howard, R: Genetic Vaccine Vector Engineering.
- WO9941383; Filed 19990210, A1 Published 19990819. Punnonen J, Bass, SH, Whalen, RG, Howard, R, Stemmer, WP: Antigen Library Immunization.
- WO9941402; Filed 19990210, A2 Published 19990819. Punnonen J, Stemmer, WP, Howard R, Patten PA: Targeting of Genetic Vaccine Vectors.
- Wong CH, Whitesides GM: Enzymes in Synthetic Organic Chemistry. Vol. 12. New York: Elsevier Science Publications, 1995.
- Yang X, Hubbard EJ, Carlson M: A protein kinase substrate identified by the two-hybrid system. *Science* 257(5070):680-2, (Jul 31) 1992.

**WHAT IS CLAIMED IS:**

1. A method for producing a library of nucleic acids encoding a plurality of modified antigen binding sites, wherein the modified antigen binding sites are derived from a first nucleic acid comprising a sequence encoding a first antigen binding site, the method  
5 comprising:

(a) providing a first nucleic acid encoding a first antigen binding site;

(b) providing a set of mutagenic oligonucleotides that encode naturally-occurring amino acid variants at a plurality of targeted codons in the first nucleic acid; and,

(c) using the set of mutagenic oligonucleotides to generate a set of antigen  
10 binding site-encoding variant nucleic acids encoding a range of amino acid variations at each amino acid codon that was mutagenized,

thereby producing a library of nucleic acids encoding a plurality of modified antigen binding sites.

15 2. The method of claim 1, wherein step (b) provides a set of mutagenic oligonucleotides that encode all nineteen naturally-occurring amino acid variants for each targeted codon, thereby generating all 19 possible natural amino acid changes at each amino acid codon mutagenized.

20 3. The method of claim 1, further comprising expressing the set of variant antigen binding site-encoding nucleic acids such that antigen binding site-encoding polypeptides encoded by the variant nucleic acids are expressed.

25 4. The method of claim 1, wherein the set of mutagenic oligonucleotides comprises a 19-fold degenerate mutagenic oligonucleotide for each codon to be mutagenized, wherein each of the 19-fold degenerate mutagenic oligonucleotides comprises a homologous first sequence and a degenerate triplet second sequence.

30 5. The method of claim 1, wherein the antigen binding site comprises a single stranded antigen binding polypeptide, a Fab fragment, an Fc fragment, a Fd fragment, a F(ab')<sub>2</sub> fragment, a Fv fragment or a complementarity determining region (CDR).

6. The method of claim 5, wherein the antigen binding site polypeptide further comprises an antibody polypeptide.

5 7. The method of claim 1, wherein the antigen binding site polypeptide further comprises an antigen binding site of a T cell receptor (TCR).

8. The method of claim 7, wherein the antigen binding site polypeptide further comprises a T cell receptor (TCR).

10 9. The method of claim 1, wherein the antigen binding site polypeptide further comprises an antigen binding site of a major histocompatibility complex (MHC) molecule.

10. The method of claim 9, wherein the antigen binding site polypeptide further comprises a major histocompatibility complex (MHC) molecule.

15 11. The method of claim 10, wherein the major histocompatibility complex (MHC) molecule comprises a Class I molecule.

20 12. The method of claim 10, wherein the major histocompatibility complex (MHC) molecule comprises a Class II molecule.

13. The method of claim 1, wherein the nucleic acid of step (a) is derived from a nucleic acid encoding a mammalian polypeptide.

25 14. The method of claim 13, wherein the mammalian polypeptide comprises a human polypeptide.

30 15. The method of claim 13, wherein the mammalian polypeptide is selected from the group consisting of an antibody, a T cell receptor, a Class I MHC molecule and a Class II MHC molecule.

16. The method of claim 1, wherein the nucleic acid of step (a) is derived from a human nucleic acid encoding an antigen binding site.

17. The method of claim 16, wherein the nucleic acid of step (a) is derived from a phage comprising a human nucleic acid sequence encoding an antigen binding site, wherein the phage expresses the antigen binding site.

18. The method of claim 16, wherein the nucleic acid of step (a) is derived from a non-human mammal comprising a human nucleic acid sequence encoding an antigen binding site, wherein the non-human mammal expresses the antigen binding site.

19. The method of claim 18, wherein the non-human mammal is a transgenic non-human mammal.

20. The method of claim 19, wherein the transgenic non-human mammal is a mouse.

21. The method of claim 1, wherein at least two amino acid codons in the antigen binding site are mutagenized.

22. The method of claim 21, wherein all the amino acid codons in the antigen binding site are mutagenized.

23. The method of claim 6, wherein all the amino acid codons in the antibody polypeptide are mutagenized.

24. The method of claim 8, wherein all the amino acid codons in the T cell receptor (TCR) are mutagenized.

25. The method of claim 10, wherein all the amino acid codons in the MHC molecule are mutagenized.

26. The method of claim 1, wherein a degenerate mutagenic oligonucleotide comprises a first homologous sequence, a degenerate triplet second sequence, and a third homologous sequence.

5 27. The method of claim 1, wherein each degenerate oligonucleotide comprises a first homologous sequence, a plurality of degenerate triplets second sequences, and a third homologous sequence.

10 28. The method of claim 3, further comprising screening the expressed antigen binding site polypeptide for its ability to specifically bind an antigen.

15 29. The method of claim 28, comprising screening the expressed antigen binding site polypeptide for its ability to specifically bind an antigen capable of being specifically bound by the first antigen binding site polypeptide.

30 30. The method of claim 29, comprising identifying an antigen binding site variant by its increased antigen binding affinity or antigen binding specificity as compared to the affinity or specificity of the first antigen binding site to the antigen.

20 31. The method of claim 29, comprising identifying an antigen binding site variant by its decreased antigen binding affinity or antigen binding specificity as compared to the affinity or specificity of the first antigen binding site to the antigen.

25 32. The method of claim 1, further comprising mutagenizing the first nucleic acid of step (a) by a method comprising an optimized directed evolution system.

33. The method of claim 1, further comprising mutagenizing the first nucleic acid of step (a) by a method comprising a synthetic ligation reassembly.

30 34. The method of claim 3, comprising screening the expressed antigen binding site polypeptide for its ability to specifically bind an antigen by a method comprising expression of the expressed antigen binding site polypeptide in a solid phase.

35. The method of claim 34, comprising screening the expressed antigen binding site polypeptide for its ability to specifically bind an antigen by a method comprising a capillary array.

5

36. The method of claim 34, comprising screening the expressed antigen binding site polypeptide for its ability to specifically bind an antigen by a method comprising a double-orificed container.

10

37. The method of claim 36, wherein the double-orificed container comprises a double-orificed capillary array.

38. The method of claim 37, wherein the double-orificed capillary array is a GIGAMATRIX™ capillary array.

15

39. The method of claim 34, comprising screening the expressed antigen binding site polypeptide for its ability to specifically bind an antigen by a method comprising use of an ELISA.

20

40. The method of claim 3, comprising screening the expressed antigen binding site polypeptide for its ability to specifically bind an antigen by a method comprising phage display of the antigen binding site polypeptide.

25

41. The method of claim 3, comprising screening the expressed antigen binding site polypeptide for its ability to specifically bind an antigen by a method comprising expression of the expressed antigen binding site polypeptide in a liquid phase.

30

42. The method of claim 3, comprising screening the expressed antigen binding site polypeptide for its ability to specifically bind an antigen by a method comprising ribosome display of the antigen binding site polypeptide.

43. The method of claim 1, wherein the set of progeny antigen binding site-encoding variant nucleic acids is generated by amplifying the nucleic acid of step (a) by a polymerase-based amplification using a plurality of oligonucleotides.

5 44. The method of claim 43, wherein the amplification comprises a polymerase chain reaction (PCR).

45. A library of nucleic acids encoding a plurality of modified antigen binding sites, wherein the modified antigen binding sites are derived from a first nucleic acid comprising a sequence encoding a first antigen binding site, made by a method comprising the following steps:

- 10 (a) providing a first nucleic acid encoding a first antigen binding site;  
(b) providing a set of mutagenic oligonucleotides that encode naturally-occurring amino acid variants at a plurality of targeted codons in the first nucleic acid; and,  
15 (c) using the set of mutagenic oligonucleotides to generate a set of antigen binding site-encoding variant nucleic acids encoding a range of amino acid variations at each amino acid codon that was mutagenized,

thereby producing a library of nucleic acids encoding a plurality of modified antigen binding sites.

20 46. A method for producing from a library of variant antibodies from a template antibody, the method comprising:

- 25 (a) providing a first nucleic acid encoding the template antibody;  
(b) providing a set of mutagenic oligonucleotides that encode naturally-occurring amino acid variants at a plurality of targeted codons in the first nucleic acid; and,  
c) using the set of mutagenic oligonucleotides to generate a set of antibody-encoding variant nucleic acids encoding a range of amino acid variations at each amino acid codon that was mutagenized,

30 thereby producing a library of nucleic acids encoding a plurality of variant antibodies.

47. The method of claim 46, wherein step (b) provides a set of mutagenic oligonucleotides that encode all nineteen naturally-occurring amino acid variants for each targeted codon, thereby generating all 19 possible natural amino acid changes at each amino acid codon mutagenized.

5

48. The method of claim 46, wherein the antibody is selected from the group consisting of polypeptides comprising a Fab fragment, an Fd fragment, an Fc fragment, a F(ab')<sub>2</sub> fragment, a Fv fragment and a complementarity determining region (CDR).

10

49. The method of claim 46, wherein the plurality of oligonucleotides comprises a degenerate oligonucleotide for each codon to be mutagenized, wherein each of the degenerate oligonucleotides comprises a homologous first sequence and a degenerate triplet second sequence.

15

50. The method of claim 46, wherein the set of progeny polynucleotides encoding antibodies is generated by amplifying the nucleic acid of step (a) using a plurality of oligonucleotides.

20

51. A library of variant antibodies derived from a template antibody made by a method comprising the following steps:

(a) providing a first nucleic acid encoding the template antibody;

(b) providing a set of mutagenic oligonucleotides that encode naturally-occurring amino acid variants at a plurality of targeted codons in the first nucleic acid; and,

25

(c) using the set of mutagenic oligonucleotides to generate a set of antibody-encoding variant nucleic acids encoding a range of amino acid variations at each amino acid codon that was mutagenized,

thereby producing a library of nucleic acids encoding a plurality of variant antibodies.

30

52. A method for producing from a library of variant T cell receptors (TCRs) from a template T cell receptor (TCR), the method comprising:

(a) providing a first nucleic acid encoding the template T cell receptor;



(b) providing a set of mutagenic oligonucleotides that encode naturally-occurring amino acid variants at a plurality of targeted codons in the first nucleic acid; and,  
c) using the set of mutagenic oligonucleotides to generate a set of T cell receptor (TCR)-encoding variant nucleic acids encoding a range of amino acid variations at  
5 each amino acid codon that was mutagenized,  
thereby producing a library of nucleic acids encoding a plurality of variant T cell receptors (TCRs).

53. A library of variant T cell receptors (TCRs) derived from a template T cell receptor (TCR) made by a method comprising the following steps:

(a) providing a first nucleic acid encoding the template T cell receptor;  
(b) providing a set of mutagenic oligonucleotides that encode naturally-occurring amino acid variants at a plurality of targeted codons in the first nucleic acid; and,  
c) using the set of mutagenic oligonucleotides to generate a set of T cell  
15 receptor (TCR)-encoding variant nucleic acids encoding a range of amino acid variations at each amino acid codon that was mutagenized,  
thereby producing a library of nucleic acids encoding a plurality of variant T cell receptors (TCRs).

54. A method for producing from a library of variant major histocompatibility complex (MHC) molecules from a template major histocompatibility complex (MHC) molecule, the method comprising:

(a) providing a first nucleic acid encoding the template major histocompatibility complex (MHC) molecule;  
25 (b) providing a set of mutagenic oligonucleotides that encode naturally-occurring amino acid variants at a plurality of targeted codons in the first nucleic acid; and,  
c) using the set of mutagenic oligonucleotides to generate a set of major histocompatibility complex (MHC) molecule-encoding variant nucleic acids encoding a range of amino acid variations at each amino acid codon that was mutagenized,  
30 thereby producing a library of nucleic acids encoding a plurality of variant major histocompatibility complex (MHC) molecules.

55. A library of variant major histocompatibility complex (MHC) molecules derived from a template major histocompatibility complex (MHC) molecule made by a method comprising the following steps:

- 5 (a) providing a first nucleic acid encoding the template major histocompatibility complex (MHC) molecule;
- (b) providing a set of mutagenic oligonucleotides that encode naturally-occurring amino acid variants at a plurality of targeted codons in the first nucleic acid; and,
- 10 (c) using the set of mutagenic oligonucleotides to generate a set of major histocompatibility complex (MHC) molecule-encoding variant nucleic acids encoding a range of amino acid variations at each amino acid codon that was mutagenized,
- thereby producing a library of nucleic acids encoding a plurality of variant major histocompatibility complex (MHC) molecules.

56. A method of making a set of nucleic acids encoding a set of antigen binding site variants comprising the steps of:

- 15 (a) providing a template nucleic acid encoding an antigen-binding polypeptide;
- (b) providing a plurality of oligonucleotides that encode all nineteen naturally-occurring amino acid variants at a single amino acid residue of the antigen-binding polypeptide; and,
- 20 (c) generating a set of progeny antigen binding site-encoding variant nucleic acids encoding a non-stochastic range of single amino acid substitutions at each amino acid codon that was mutagenized, whereby all 19 possible natural amino acid changes are generated at each amino acid codon mutagenized,
- 25 thereby making a set of nucleic acids encoding a set of antigen binding site variants.

57. The method of claim 56, further comprising expressing the set of progeny antigen binding site-encoding polynucleotides such that antigen binding site-encoding polypeptides encoded by the progeny polynucleotides are expressed.

30

58. The method of claim 56, wherein the plurality of oligonucleotides comprises a set of degenerate oligonucleotides and each of the degenerate oligonucleotides comprises a homologous first sequence and a degenerate triplet second sequence.

5 59. The method of claim 56, wherein the antigen binding site-encoding polypeptide comprises a single stranded antigen binding polypeptide.

60. The method of claim 56, wherein the antigen binding site-encoding polypeptide comprises an antibody polypeptide.

10 61. The method of claim 56, wherein the antigen binding site-encoding polypeptide comprises an antigen binding site of a T cell receptor (TCR).

15 62. The method of claim 61, wherein the antigen binding site-encoding polypeptide further comprises a T cell receptor (TCR).

63. The method of claim 56, wherein the antigen binding site-encoding polypeptide comprises an antigen binding site of a major histocompatibility complex (MHC) molecule.

20 64. The method of claim 63, wherein the antigen binding site-encoding polypeptide further comprises a major histocompatibility complex (MHC) molecule.

25 65. The method of claim 56, wherein the nucleic acid of step (a) is derived from a nucleic acid encoding a mammalian antibody polypeptide.

66. The method of claim 65, wherein the nucleic acid of step (a) is derived from a human nucleic acid.

30 67. The method of claim 56, wherein at least two amino acid codons in the antigen binding site are mutagenized and a set of degenerate oligonucleotides that encode all

nineteen naturally-occurring amino acid variants are provided for each amino acid codon mutagenized.

68. The method of claim 56, wherein all the amino acid codons in the antigen binding site are mutagenized and a set of degenerate oligonucleotides that encode all nineteen naturally-occurring amino acid variants are provided for each amino acid codon mutagenized.

69. The method of claim 60, wherein all the amino acid codons in the antibody polypeptide are mutagenized.

70. The method of claim 61, wherein all the amino acid codons in the antigen binding site of the T cell receptor (TCR) are mutagenized.

71. The method of claim 63, wherein all the amino acid codons in the antigen binding site of the major histocompatibility complex (MHC) molecule are mutagenized.

72. The method of claim 56, wherein a degenerate oligonucleotide comprises a first homologous sequence, a degenerate triplet second sequence, and a homologous third sequence.

73. The method of claim 56, wherein each degenerate oligonucleotide comprises a first homologous sequence, a degenerate triplet second sequence, and a homologous third sequence.

74. The method of claim 57, further comprising screening an expressed antigen binding site-encoding polypeptide for its ability to specifically bind an antigen.

75. The method of claim 57, comprising screening the expressed antigen binding site-encoding polypeptide for its ability to specifically bind an antigen capable of being specifically bound by the first antigen binding site.

76. The method of claim 75, comprising identifying an antigen binding site variant by its increased antigen binding affinity or antigen binding specificity to the antigen as compared to the affinity or specificity of the antigen binding site encoded by the nucleic acid of step (a).

5

77. The method of claim 56, further comprising mutagenizing the template nucleic acid by a method comprising an optimized directed evolution system.

78. The method of claim 56, further comprising mutagenizing the template nucleic acid by a method comprising a synthetic ligation reassembly.

10

79. The method of claim 56, comprising screening the expressed antigen binding site-encoding polypeptide for its ability to specifically bind an antigen by a method comprising a capillary array.

15

80. The method of claim 56, comprising screening the expressed antigen binding site-encoding polypeptide for its ability to specifically bind an antigen by an ELISA.

81. The method of claim 56, wherein the set of variant nucleic acids is generated by performing amplification reactions on the nucleic acid of step (a) using the set of oligonucleotides to generate a set of variant nucleic acids encoding nineteen amino acid substitution variants at a single amino acid residue of the antigen-binding polypeptide.

20

82. The method of claim 81, wherein the amplification comprises a polymerase-based amplification.

25

83. The method of claim 82, wherein polymerase-based amplification comprises a polymerase chain reaction (PCR).

84. The method of claim 56, wherein the set of variant nucleic acids comprises  $10^{10}$  members.

30

85. The method of claim 56, wherein the set of variant nucleic acids comprises  $10^5$  members.

86. The method of claim 56, wherein the set of variant nucleic acids comprises  $10^3$  members.

87. A method of making a set of antibody variants comprising the steps of:

- (a) providing a nucleic acid encoding an antibody;
- (b) providing a plurality of oligonucleotides;
- (c) generating a non-stochastic range of single amino acid substitutions at each amino acid codon, whereby all 19 possible natural amino acid changes are generated at each amino acid codon mutagenized, thereby generating a set of variant nucleic acids; and,
- (d) expressing the set of variant nucleic acids such that the antibody variants encoded by the variant nucleic acids are expressed.

88. The method of claim 87, wherein the antibody is selected from the group consisting of polypeptides comprising a Fab fragment, a Fd fragment, an Fc fragment, a F(ab')<sub>2</sub> fragment, a Fv fragment and a complementarity determining region (CDR).

89. The method of claim 87, wherein the plurality of oligonucleotides comprises a set of degenerate oligonucleotides that encode all nineteen naturally-occurring amino acid variants at a single amino acid residue of the antibody, wherein each of the degenerate oligonucleotides comprises a homologous first sequence and a degenerate triplet second sequence.

90. The method of claim 87, wherein generating a non-stochastic range of single amino acid substitutions comprises performing amplification reactions on the nucleic acid of step (a) using the set of oligonucleotides to generate a set of variant nucleic acids encoding nineteen amino acid substitution variants at a single amino acid residue of the antibody.

91. A method of identifying a variant of an antigen binding site comprising the steps of:

- (a) providing a nucleic acid encoding an antigen binding site;
- (b) providing a set of oligonucleotides that encode all nineteen naturally-occurring amino acid variants at all residues of the antigen-binding site;
- (c) incorporating the sequence of the oligonucleotides of step (b) into the nucleic acid of step (a) to generate a set of variant nucleic acids encoding nineteen amino acid substitution variants at each residue of the antigen binding site;
- (d) expressing each of the variant nucleic acids as polypeptides and measuring the variant's affinity to the antigen; and,
- (e) identifying a variant of the antigen binding site by its increased or decreased antigen binding specificity as compared to the antigen binding affinity of the antigen binding site encoded by the nucleic acid of step (a).

92. The method of claim 91, wherein the variant nucleic acids are expressed using *in vitro* transcription/translation.

93. The method of claim 91, wherein the variant nucleic acids are expressed using phage display.

94. The method of claim 91, wherein the variant nucleic acids are expressed using ribosome display.

95. The method of claim 91, wherein the variant nucleic acids are expressed using a double orificed container.

96. The method of claim 95, wherein the variant nucleic acids are expressed using a double orificed capillary array.

97. The method of claim 91, wherein the set of oligonucleotides comprises a set of degenerate oligonucleotides that encode all nineteen naturally-occurring amino acid variants at a single amino acid residue of the antibody, wherein each of the degenerate oligonucleotides comprises a homologous first sequence and a degenerate triplet second sequence.

98. The method of claim 91, wherein the antigen binding site comprises an antibody.

5 99. The method of claim 98, wherein the antibody is selected from the group consisting of polypeptides comprising a Fab fragment, an Fd fragment, an Fc fragment, a F(ab')<sub>2</sub> fragment, a Fv fragment and a complementarity determining region (CDR).

10 100. The method of claim 91, wherein the antigen binding site comprises an antigen binding site of a T cell receptor.

101. The method of claim 91, wherein the antigen binding site comprises an antigen binding site of a major histocompatibility complex molecule.

15 102. The method of claim 91, wherein incorporating the sequence of the oligonucleotides of step (b) into the nucleic acid of step (a) is accomplished by an amplification reaction using the oligonucleotides as primers.



# Exo III Generated Structures

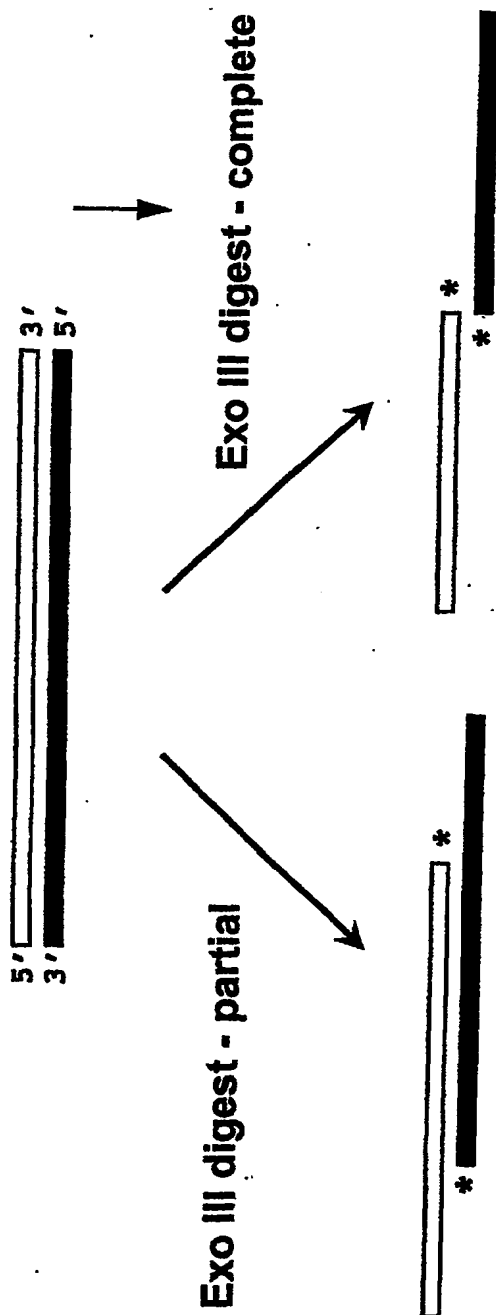
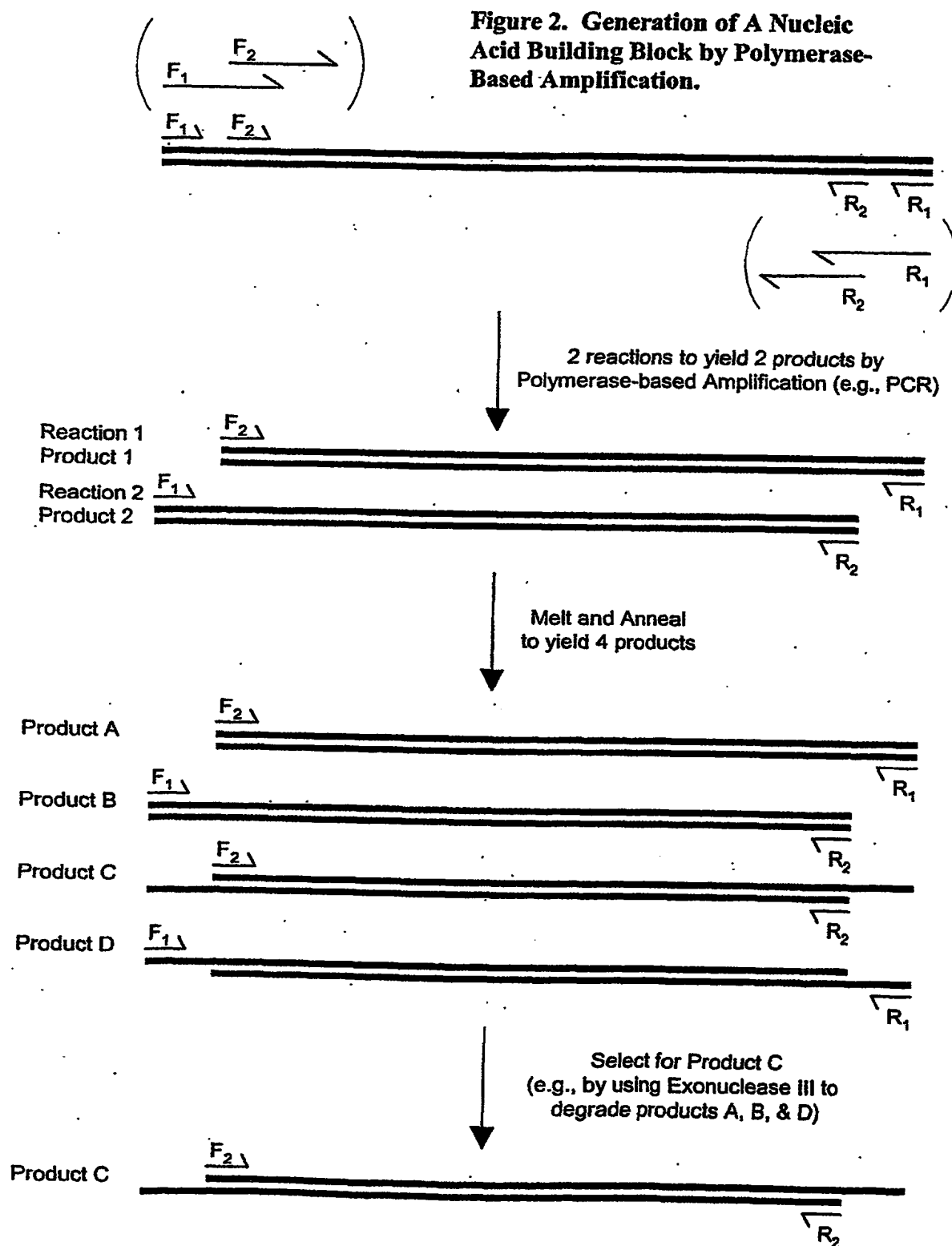


Figure 1



### FIGURE 3. Unique Overhangs And Unique Couplings.

The number of unique overhangs of each size (e.g. the total number of unique overhangs composed of 1 or 2 or 3, etc. nucleotides) exceeds the number of unique couplings that can result from the use of all the unique overhangs of that size. For example, the total number of unique couplings that can be made using all the 8 unique single-nucleotide 3' overhangs and single-nucleotide 5' overhangs is 4.

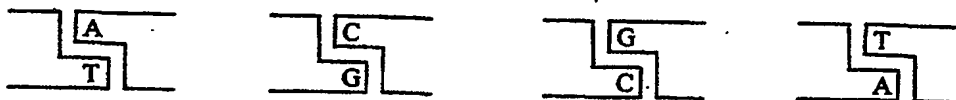
**PANEL A.** 4 unique single-nucleotide 3' overhangs are possible (i.e., A, C, G, & T). For each of these there is a complementary 3' overhang with which it can pair (i.e., T, G, C, & A, respectively), as shown.



**PANEL B.** However, the number of unique single-nucleotide 3' overhangs is greater than the number of unique couplings. Thus, only 2 intrinsically unique couplings exist using single-nucleotide 3' overhangs as shown.



**PANEL C.** Likewise, 4 unique-single nucleotide 5' overhangs are possible (i.e., A, C, G, & T). For each of these there is a complementary 5' overhang with which it can pair (i.e., T, G, C, & A, respectively), as shown.



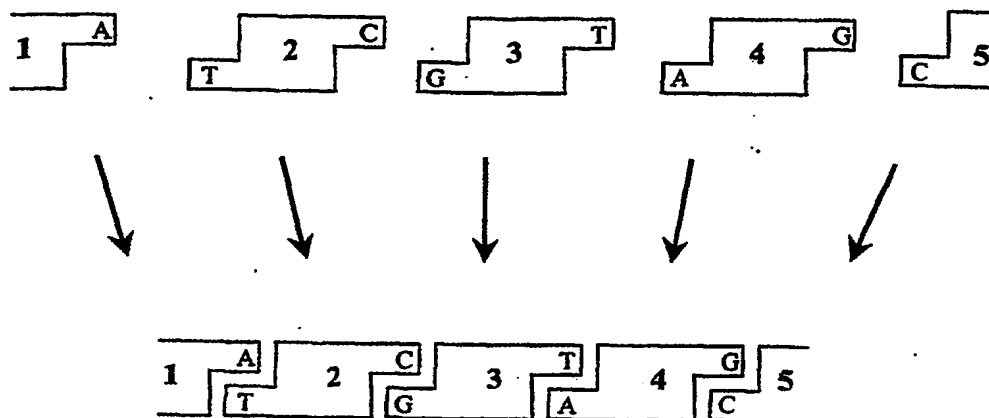
**PANEL D.** However, the number of unique single-nucleotide 5' overhangs is greater than the number of unique couplings. Thus, only 2 intrinsically unique couplings exist using single-nucleotide 5' overhangs as shown.



# FIGURE 4. Unique Overall Assembly Order Achieved by Sequentially Coupling the Building Blocks

Awareness of the degeneracy (between the number of unique overhangs and the number of unique couplings) is important in order to avoid the production of degeneracy in the overall assembly order of the finalized nucleic acid. However, a unique overall assembly order can also be achieved - despite the use of non-unique couplings - by using building blocks having distinct combinations of couplings, and/or by stepping the assembly of the building blocks in a deliberately chosen sequence.

**PANEL A.** For example, one could attempt to assemble the following nucleic acid product using the 5 nucleic acid building blocks as shown.

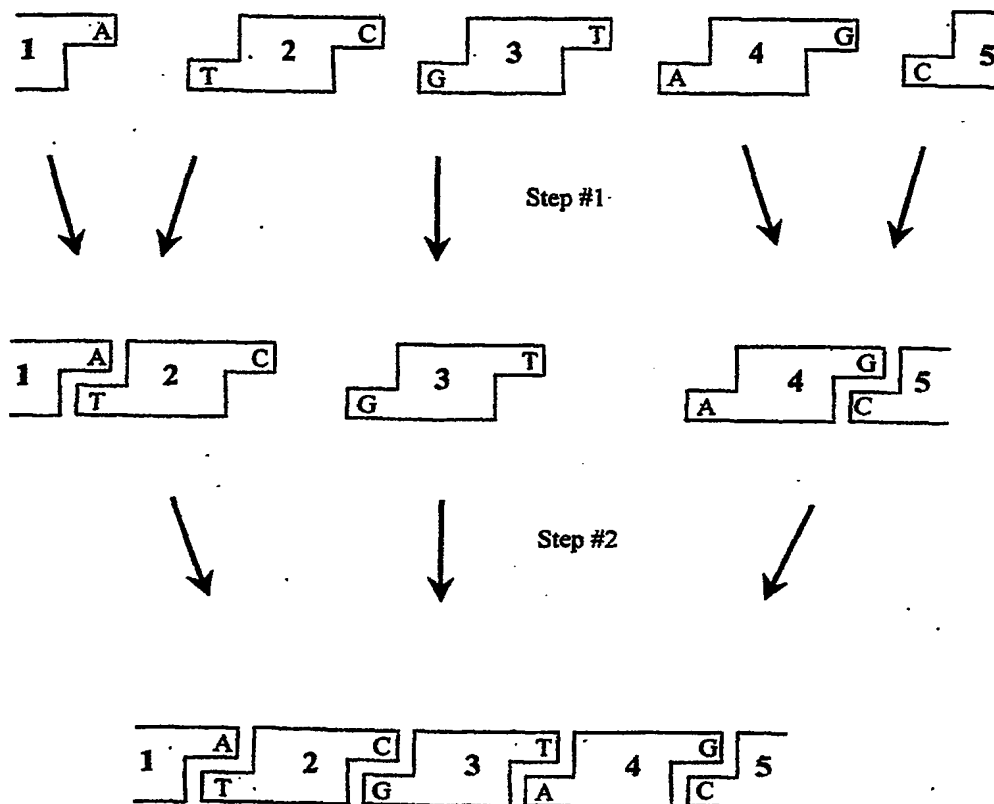


**PANEL B.** However, degeneracy in the overall assembly order of the 5 nucleic acid building blocks would be present if the assembly process were carried out in one step. For example, building block #2 and building block #3 could both couple to building block #1 as shown.



**FIGURE 4 cont.**

**PANEL C.** However, a unique overall assembly order could be achieved by sequentially coupling the building blocks in 2 steps (rather than all at once) as shown.



**Figure 5. Unique Couplings Available Using a Two-Nucleotide 3' Overhang.**

16 unique 3' overhangs can be formed using two-nucleotides. However, use of these 16 unique overhangs allows for the formation of only 6 unique couplings. Another 6 unique couplings are provided by the use 5' overhangs formed using two-nucleotides. Thus, a total of 12 unique couplings are provided by the combined use of 3' and 5' two-nucleotide overhangs. "Twin" couplings are marked in the same shading.

		TOP STRAND 2 <sup>ND</sup> Overhanging Nucleotide (counting from 5' to 3')					
		A	C	G	T		
TOP STRAND 1 <sup>ST</sup> Overhanging Nucleotide (counting from 5' to 3')	A	AA TT	AC TG	AG TC	PALINDROMIC AT TA	BOTTOM STRAND 2 <sup>ND</sup> Overhanging Nucleotide (counting from 5' to 3')	T
	C	CA GT	CC GG	PALINDROMIC CG GC	CT GA		G
	G	GA CT	PALINDROMIC GC CG	GG CC	GT CA		C
	T	PALINDROMIC TA AT	TC AG	TG AC	TT AA		A
		T	G	C	A		
		BOTTOM STRAND 1 <sup>ST</sup> Overhanging Nucleotide (counting from 5' to 3')					

**Figure 6. Generation of an Exhaustive Set of Chimeric Combinations by Synthetic Ligation Reassembly.**

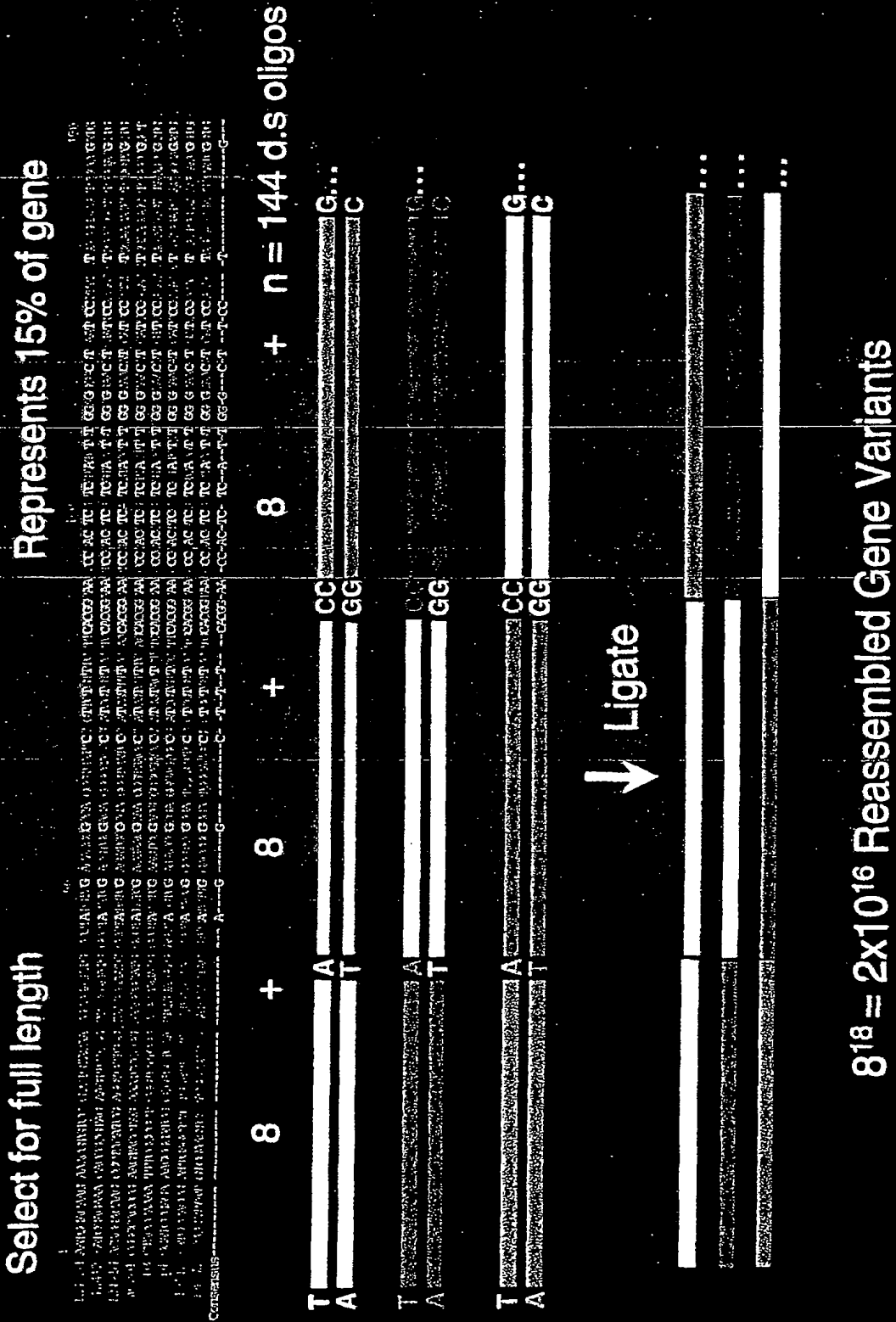


Figure 6. Unique Couplings Available Using a Three-Nucleotide Overhang.

		TOP STRAND - 1 <sup>ST</sup> Overhanging Nucleotide (BOTTOM STRAND – Complementary Nucleotide)					
		A	C	G	T		
TOP STRAND – 2 <sup>nd</sup> Overhanging Nucleotide (BOTTOM STRAND – Complementary Nucleotide)	A	AAA	CAA	GAA	TAA	A	TOP STRAND – 3 <sup>rd</sup> Overhanging Nucleotide (BOTTOM STRAND – Complementary Nucleotide)
		AAC	CAC	GAC	TAC	C	
		AAG	CAG	GAG	TAG	G	
		AAT	CAT	GAT	TAT	T	
	C	ACA	CCA	GCA	TCA	A	
		ACC	CCC	GCC	TCC	C	
		ACG	CCG	GCG	TCG	G	
		ACT	CCT	GCT	TCT	T	
	G	AGA	CGA	GGA	TGA	A	
		AGC	CGC	GGC	TGC	C	
		AGG	CGG	GGG	TGG	G	
		AGT	CGT	GGT	TGT	T	
	T	ATA	CTA	GTA	TTA	A	
		ATC	CTC	GTC	TTC	C	
		ATG	CTG	GTG	TTG	G	
		ATT	CTT	GTT	TTT	T	
		T	G	C	A		
		BOTTOM STRAND 1 <sup>ST</sup> Overhanging Nucleotide (counting from 5' to 3')					



Figure 6. Unique Couplings Available Using a Three-Nucleotide 3' Overhang.

1 <sup>st</sup> Base	TOP STRAND			BOTTOM STRAND		No.	Comments
	2 <sup>nd</sup> Base	3 <sup>rd</sup> Base	Sequence 5'-XXX-3'	Sequence 3'-XXX-5'	Sequence 5'-XXX-3'		
A	A	A	AAA	TTT	TTT	1	
		C	AAC	TTG	GTT	2	
		G	AAG	TTC	CTT	3	
		T	AAT	TTA	ATT	4	
	C	A	ACA	TGT	TGT	5	
		C	ACC	TGG	GGT	6	
		G	ACG	TGC	CGT	7	
		T	ACT	TGA	AGT	8	
	G	A	AGA	TCT	TCT	9	
		C	AGC	TCG	GCT	10	
		G	AGG	TCC	CCT	11	
		T	AGT	TCA	ACT	12	
	T	A	ATA	TAT	TAT	13	
		C	ATC	TAG	GAT	14	
		G	ATG	TAC	CAT	15	
		T	ATT	TAA	AAT	16	
C	A	A	CAA	GTT	TTG	17	
		C	CAC	GTG	GTG	18	
		G	CAG	GTC	CTG	19	
		T	CAT	GTA	ATG	20	
	C	A	CCA	GGT	TGG	21	
		C	CCC	GGG	GGG	22	
		G	CCG	GGC	CGG	23	
		T	CCT	GGA	AGG	24	
	G	A	CGA	GCT	TCG	25	
		C	CGC	GCG		26	
		G	CGG	GCC		27	
		T	CGT	GCA		28	
	T	A	CTA	GAT		29	
		C	CTC	GAG		30	
		G	CTG	GAC		31	
		T	CTT	GAA		32	
G	A	A	GAA	CTT		33	
		C	GAC	CTG		34	
		G	GAG	CTC		35	
		T	GAT	CTA		36	
	C	A	GCA	CGT		37	
		C	GCC	CGG		38	
		G	GCG	CGC		39	
		T	GCT	CGA		40	
	G	A	GGA	CCT		41	
		C	GGC	CCG		42	
		G	GGG	CCC		43	
		T	GGT	CCA		44	
	T	A	GTA	CAT		45	
		C	GTC	CAG		46	
		G	GTG	CAC		47	
		T	GTT	CAA		48	
T	A	A	TAA	ATT		49	
		C	TAC	ATG		50	
		G	TAG	ATC		51	
		T	TAT	ATA		52	
	C	A	TCA	AGT		53	
		C	TCC	AGG		54	
		G	TCG	AGC		55	
		T	TCT	AGA		56	
	G	A	TGA	ACT		57	
		C	TGC	ACG		58	
		G	TGG	ACC		59	
		T	TGT	ACA		60	
	T	A	TTA	AAT		61	
		C	TTC	AAG		62	
		G	TTG	AAC		63	
		T	TTT	AAA		64	

Figure 7. Synthetic genes from oligos.

150am13_00	NcoI	c	ATGATGCACG	GCGATATTTT	ATCGAGCAAT	GACACGGTCG	GCGTTGCCGT	CCGT
150AM7_001		c	ATGCATCACG	GCGACATTTT	ATCGAGCAAT	GACACGGTCG	GCGTTGCCGT	CCGT
431am7_002		c	ATGAGACACG	GAGATATCTC	CAGCAGCAAC	GATTGCGTGG	GCGTGGCCGT	CCGT
150am13_00			CGTGAACCTAC	AAGATGCCTC	GCCTTCATAC	CAAGGCCGAG	GTTTTAGCGA	GAG GT
150AM7_001			CGTGAACCTAC	AAGATGCCGC	GGCTTCACAC	CAAGGCTGAG	GTGCTGGCCA	
431am7_002			CGTGAACCTAC	AAGATGCCGC	GGCTGCATAC	CCGCGCGGAG	GTGATGGAGA	
150am13_00			ACGCCAGAAA	GATCGGCGAG	ATGATCGTCG	GCATGAAGAC	CGGCCTGCCC	CGG
150AM7_001			ACTGCCGCAA	GATCGCCGAC	ATGCTGGTCG	GCATGAAGAG	CGGCCTGCCC	
431am7_002			ACGCCCCGAA	GATCGCCGAC	ATGGTCGTGG	GCATGAAGCG	CGGCCTGCCC	
150am13_00			GGAATGGATC	TGGTGATCTT	CCCGGAATAT	TCGACCCACG	GCATCATGTA	CCACG
150AM7_001			GGAATGGATC	TGGTGATCTT	CCCGGAATAT	TCCACCCACG	GCATCATGTA	
431am7_002			GGCATGGACC	TGGTCATCTT	CCCCGAGTAC	TCCACCCACG	GCATCATGTA	
150am13_00			CGACTCCAAG	GAAATGTACG	ATACCGCGTC	CGTCGTGCCC	GGCGAGGAGA	CCC GG
150AM7_001			CGACTCCAAG	GAGATGTACG	ACACGGCGTC	GACGGTGCCG	GGTGAAGAGA	
431am7_002			CGACGCCAAG	GAAATGTACG	AAACCGCTTC	GGCCATGCCG	GGCGAAGAGA	
150am13_00			CCGAGATTTT	TGCCGAAGCC	TGCCGCAAGG	CGAAAGTCTG	GGGCGTGTTT	G GGG
150AM7_001			CCGAGATTTT	CGCCGAGGCC	TGCCGCAAGG	CCAAGGTCTG	GGGCGTGTTT	
431am7_002			CTGCTGTGTT	CGCCGACGCC	TGCCGCAAGG	CCAACGTATG	GGGCGTGTTT	
150am13_00			TCGCTCACCG	GCGAACGTCA	CGAGGAACAT	CCGAAGAAGG	CGCCCTACAA	AAAG C
150AM7_001			TCGCTGACCG	GCGAGCGCCA	CGAGGAGCAT	CCCAATAAAG	CGCCGTACAA	
431am7_002			TCGCTGACCG	GCGAGCGCCA	CGAAGAGCAC	CCGAACAAGG	CGCCGTACAA	
150am13_00			CACGCTGATC	CTGATGAACG	ACAAGGGCGA	GGTGGTCCAG	AAATACCGCA	CAG AA
150AM7_001			CACCTGATC	CTGATGAACG	ACAAGGGTGA	AGTCGTTCCAG	AAATATCGCA	
431am7_002			CACGCTCATC	CTGATGAACA	ACAAGGGCGA	GATCGTCCAG	AAATACCGCA	
150am13_00			AGATCATGCC	GTGGGTTCGG	ATCGAGGGCT	GGTATCCCCGG	CAACTGCACC	GGTA
150AM7_001			AGATCATGCC	GTGGGTGCCG	ATCGAAGGCT	GGTATCCCCGG	CAACTGCACC	
431am7_002			AGATCATGCC	CTGGGTGCCG	ATCGAAGGCT	GGTATCCCCGG	CGATTGCACC	
150am13_00			TACGTCTCCG	ACGGGCCGAA	GGGCAATGAAG	GTTTCGCTGA	TCATCTGCGA	TGAAG
150AM7_001			TACGTCTCCG	AAGGCCCGAA	GGGCAATGAAG	ATGTCGCTGA	TCATCTGCGA	
431am7_002			TATGTGTCGG	AAGGCCCGAA	GGGCAATGAAG	ATCAGCCTCA	TCATCTGCGA	
150am13_00			TGACGGCAAC	TATCCGGAAA	TCTGGCGCGA	CTGCGCCATG	AAGGGCGCCG	TCTGGCG
150AM7_001			CGACGGCAAC	TATCCGGAAA	TCTGGCGTGA	CTGCGCGATG	AAGGGCGCCG	
431am7_002			CGACGGCAAT	TATCCCGAGA	TCTGGCGCGA	TTGCGCCATG	CGCGGCGCCG	

Figure 7 cont.

			CCAG			
150am13_00	AGCTGATCGT	GCGCTG	CCAG	GGCTACATGT	ATCCGGCCAA	GGACCAGCAG
150AM7_001	AACTGATCAT	CCGCTG	CCAG	GGCTACATGT	ATCCCGCCAA	GGATCAGCAG
431am7_002	AGCTGATCGT	GCGTTG	CCAG	GGATACATGT	ACCCGGCCAA	GGACCAGCAG

			GC			
150am13_00	GTCATCATGG	CGAAG	GCAT	GGCGTGGGCG	AATAATTGTT	ACGTCGCGGT
150AM7_001	GTGCTGATGG	CGAAAG	CAAT	GGCCTGGGCC	AACAACGTTT	ATGTCGCGGT
431am7_002	GTCATGGTGT	CCAAG	GCAT	GGCGTGGATG	AACAACGTCT	ACGTGGCGGT

			GGGCTTCG			
150am13_00	TTCCAATGCC	GCGGGCTTCG		ATGGCGTCTA	TTCTATTTC	GGCCACTCGG
150AM7_001	CGCCAATGCC	TCGGGCTTCG		ACGGCGTCTA	CTCTATTTC	GGCCATTTCG
431am7_002	GGCCAATGCC	GCGGGCTTCG		ACGGCGTGTA	TTCTACTTC	GGCCATTTCG

			TTCGA			
150am13_00	CGATCATCGG	CTTCGAT	TGGC	CGCACGCTCG	GCGAATGCGG	CGAGGAAGAA
150AM7_001	CGATCATCGG	CTTCGAC	CGGC	CGTACCTCG	GCGAATGCGG	CGAGGAGGAT
431am7_002	CCATCATCGG	CTTCGAC	CGGC	CGCACGCTGG	GCGAATGCGG	TGAAGAAGAC

			C AGTA			
150am13_00	TACGGCATCC	AGTATG	CCCA	GCTTTGGAAG	ATGCTGATCC	GCGACGCCCG
150AM7_001	TATGGCATCC	AGTATG	CCGC	CATCTCCAAG	TCGCTGATCC	GCGACGCGCG
431am7_002	ATGGGCGTGC	AGTAC	CGCCGA	GCTCTCCACC	AGCCTGATCC	GCGACGCGCG

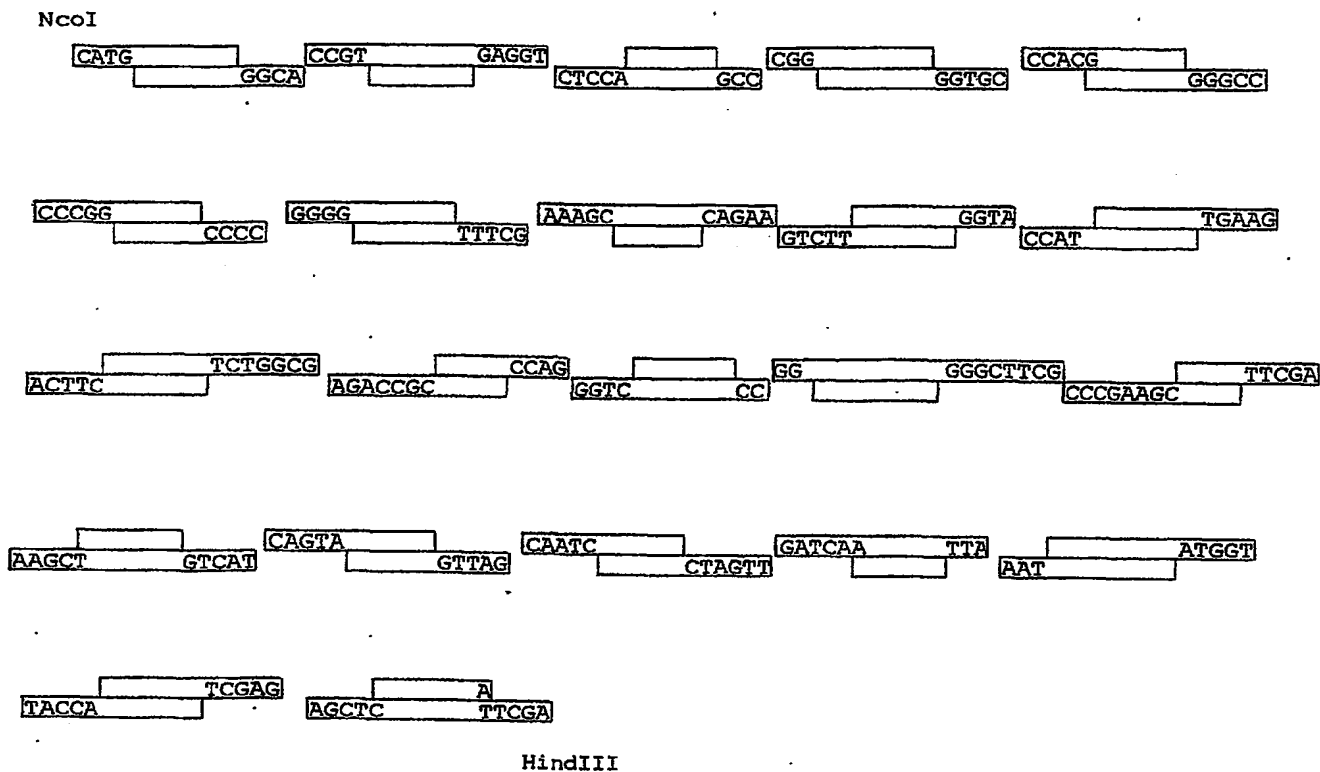
			CAATC			
150am13_00	CCGCACCGGA	CAATC	GGAAA	ACCATCTCTT	CAAGCTGGTG	CATCGTGGCT
150AM7_001	CCGCACCGGC	CAATC	GGAAA	ACCATCTCTT	CAAGCTGGTG	CACCGTGGCT
431am7_002	CAAGAACATG	CAGTC	GCAGA	ACCACTTGTT	CAAGCTGGTG	CACCGCGGCT

			GATCAA			
150am13_00	ACACCGGGTT	GATCAA	TCC	GGCGAGGGCG	ACCGCGGTCT	CGCGGCCTGT
150AM7_001	ACACCGGCAT	GATCAA	TCC	GGCGAGGGCG	ACCGCGGTGT	CGCGGCTTGC
431am7_002	ACACCGGCAA	GATCAA	TCC	GGCGAAGAGG	CCACCGGCGT	CGCGGCATGC

			TTA			
150am13_00	CGTTATGAGT	TCTACAACAA		ATGGATCGCC	GATCCGGAAG	GCACCCGCGA
150AM7_001	CGGTATGATT	TCTATTCGAA		ATGGATCGCC	GATCCCGAGG	GTACACGCGA
431am7_002	CGGTACAAC	TCTACGCCAA		CTGGATCAAC	GATCCGGAAG	GCACGCGCAA

			ATGGT			
150am13_00	ATGGT	TCGAG		TCCTTTACCC	GGCCGACGGT	GGGAACCGAT
150AM7_001	GATGGT	GGAA		TCCTTCACGC	GTCCGACGGT	GGGTGTGGAG
431am7_002	GATGGT	CGAA		TCCTTCACCC	GGTCCACCGT	GGGCACGCCG

			TCGAG			
150am13_00	TCGAAGGCAT	CCCGAACAAG		GTCGCGGTGC	ACCGCTGA	aagct
150AM7_001	TCGAGGGCAT	TCCGAACAAG		GCCACCACGC	ACCGCTGA	aagct
431am7_002	TGGACGGCAT	CCCCAACGAG		GACGCCAAGC	ACCGCTAG	aagct
						HindIII

**Figure 8. Nucleic acid building blocks for synthetic ligation gene reassembly.**

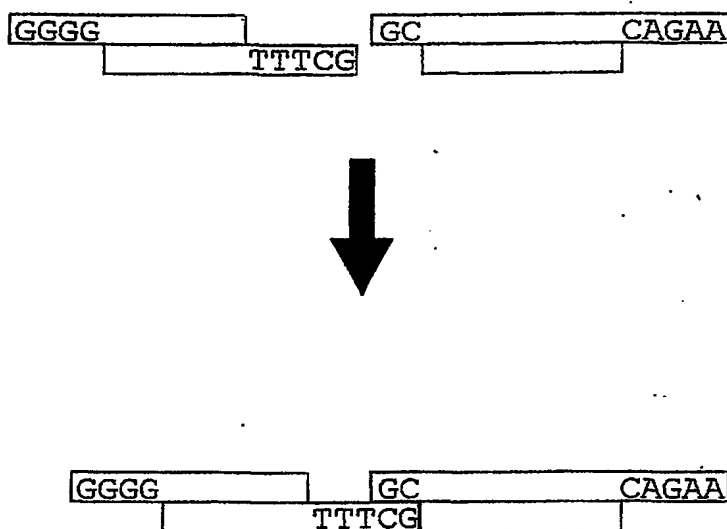
**Figure 9. Addition of Introns by Synthetic Ligation Reassembly.**

NcoI



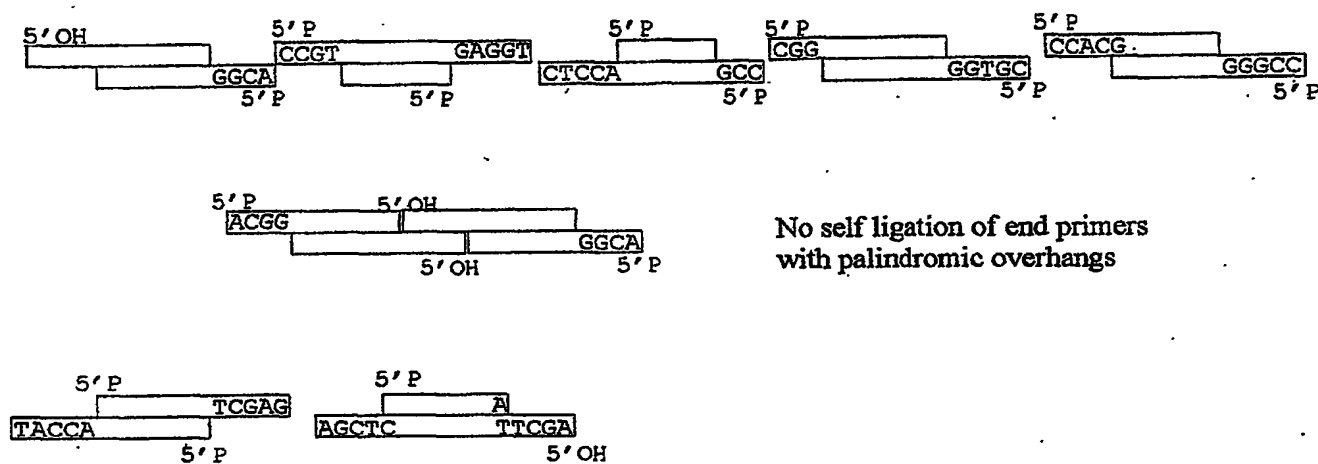
**Figure 10. Ligation Reassembly Using Fewer Than All The Nucleotides Of An Overhang.**

## Gap Ligation



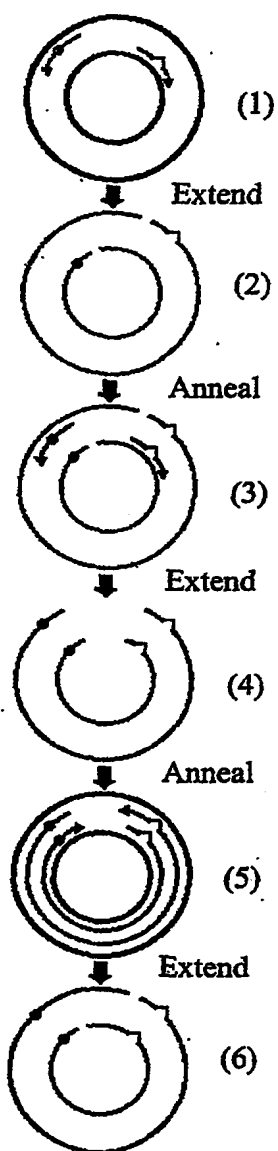
**Ligation of one strand only;  
gap in second strand can be repaired in vivo**

**Figure 11. Avoidance of unwanted self-ligation in palindromic couplings.**



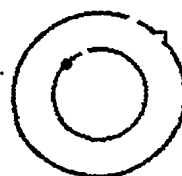
## Site-Directed Mutagenesis

**Figure 12A**



**Figure 12B**

Amplification products are comprised of the following molecular structures:



Molecule (A)

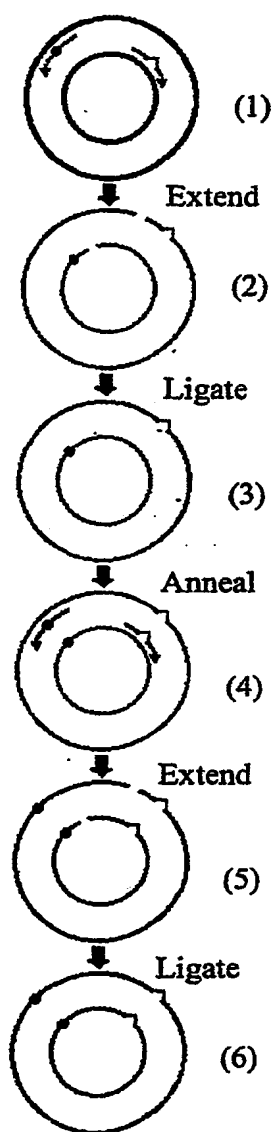


Molecule (B)



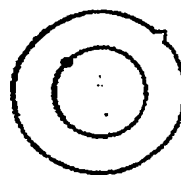
## Site-Directed Mutagenesis

**Figure 13A**

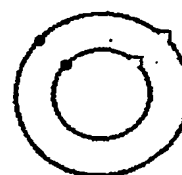


**Figure 13B**

Amplification products are comprised of the following molecular structures:



Molecule (A)

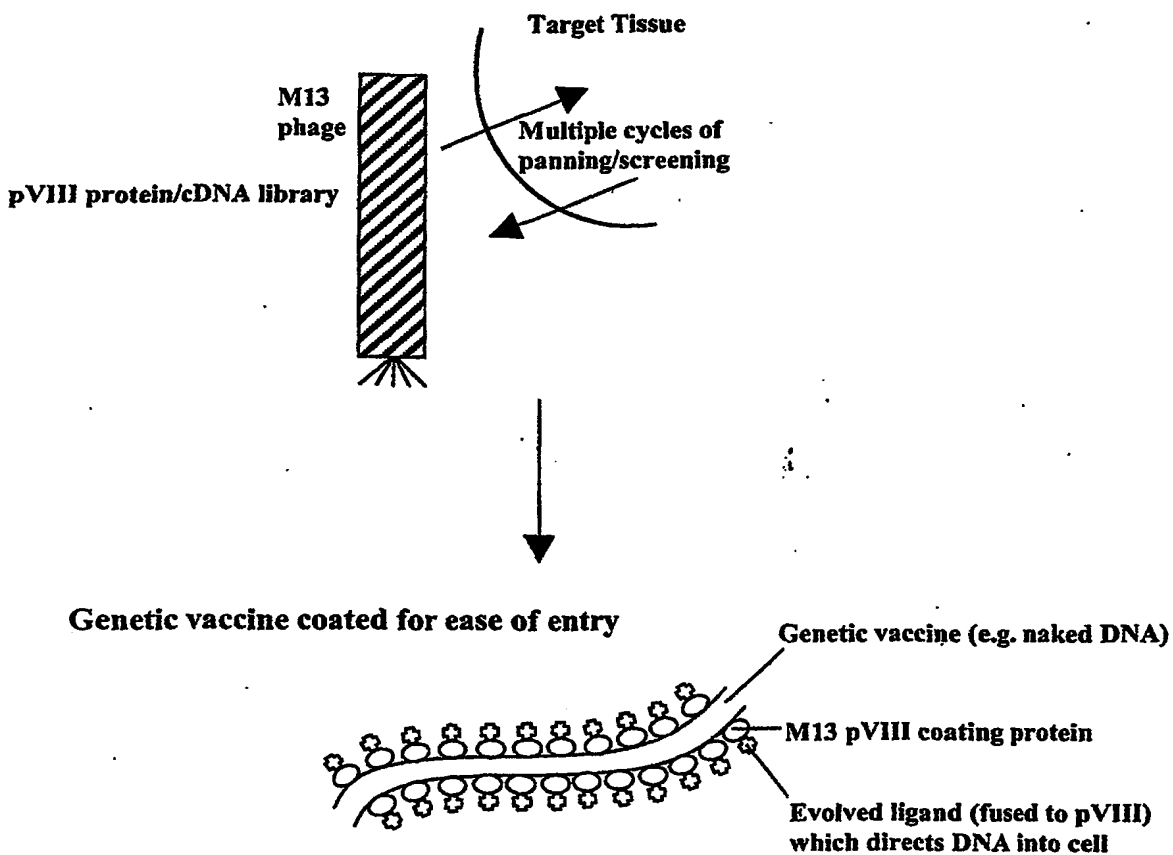


Molecule (B)

**Figure 14**

**Strategy for obtaining and using nucleic acid binding proteins that facilitate entry of genetic vaccines.**

**Evolution in M13 Format**



**Figure 15**

**A schematic representation of a method for evolving a chimeric, multivalent antigen that has immunogenic regions from multiple antigens.**

